

20. RADIOLARIANS FROM THE WESTERN NORTH PACIFIC, LEG 57, DEEP SEA DRILLING PROJECT

Richard A. Reynolds,¹ Rice University, Houston, Texas

INTRODUCTION

This is a preliminary investigation of radiolarian-bearing sediments recovered from the four DSDP Leg 57 sites drilled on the west wall of the Japan Trench. The site locations are as follows:

Site	Hole	Latitude	Longitude	Water Depth (m)	Pene-tration (m)	Oldest Sediment Cored
438	438	40°37.75'N	143°13.90'E	1552	109.5	lower Pliocene
	438A	40°37.79'N	143°14.15'E	1558	878.0	middle Miocene
	438B	40°37.80'N	143°14.80'E	1564.5	1040.7	lower Miocene
439	439	40°37.61'N	143°18.63'E	1656	1157.5	Cretaceous
440	440	39°44.13'N	143°55.74'E	4509	73.0	Pleistocene
	440A	39°44.13'N	143°55.74'E	4509	139.5	Pleistocene
	440B	39°44.13'N	143°55.74'E	4509	814.0	upper Miocene
441	441	39°45.05'N	144°04.59'E	5655	273.0	lower Pliocene
	441A	39°45.05'N	143°04.59'E	5644	662.0	upper Miocene
	441B	39°45.08'N	143°04.60'E	5635	687.0	upper Miocene

The majority of the sedimentary material recovered is Neogene and contains assemblages of radiolarians rich and diverse enough to allow biostratigraphic analysis. From such an analysis, a Neogene radiolarian biozonation is proposed for the region studied.

Paleogene and Cretaceous strata were recovered at Site 439, but the poor preservation and diversity of radiolarians inhibited analysis of these sediments. Reworking of Cretaceous radiolarians into Oligocene sediments (dated by Keller, this volume) lacking an Oligocene radiolarian fauna precluded age assignments in the lower portion of Hole 439.

Sites 438 and 440 provide excellent Neogene radiolarian sections and may prove invaluable for correlation of radiolarian events to those of other fossil groups, especially diatoms. The zonation proposed in this report is developed basically from these sections. Site 440 is especially appropriate for the recognition of Pliocene to Pleistocene zones because this interval is represented by an expanded sedimentary section. The continuous section recovered at Site 438 makes it useful for the lower Miocene to Recent interval.

SCOPE

Radiolarian biostratigraphy comprises the majority of this investigation. Preliminary radiolarian paleoecologic results are reported herein, but a more detailed analysis is required and is forthcoming (Reynolds, in preparation).

The provincialism of radiolarians has been well documented and has inhibited worldwide correlations. Materi-

als recovered on DSDP Leg 57 has afforded the opportunity to correlate a number of the "warm"-water or low-latitude radiolarian datums of Riedel and Sanfilippo (1970, 1977, 1978) to the "cold"-water or middle- or high-latitude radiolarian datums of Hays (1970), Kling (1973), and Forman (1975). This has led to the development of a Neogene radiolarian biozonation containing datums of both "warm"- and "cold"-water radiolarians. Because this is a preliminary investigation, only the most extensive and consistent datums are reported and used in the zonation.

METHODOLOGY AND PRESENTATION OF RESULTS

Samples were prepared for microscopic examination using the standard radiolarian preparation techniques of Riedel (1957) and Riedel and Sanfilippo (1977). When possible, three samples per core, with an interval of approximately three meters between them, were examined. Preliminary measurements and observations were made from these samples.

Species abundance (Tables 1-8) was determined by counting the number of individuals of a particular species per slide. Two slides were made for each sample and species categorized as follows: 1 = 1-4 individuals per slide; 2 = 5-9 individuals per slide; 3 = 10-14 individuals per slide; 4 = 15-24 individuals per slide; 5 = 25-34 individuals per slide; and 6 = more than 35 individuals per slide. A dot (•) indicates that only one isolated individual was observed in the two slides. Not all of the species observed in this study are reported herein—only those with apparent biostratigraphic importance. The most consistently abundant groups encountered are the spongodiscids, actinomiids, and litheliids.

Relative preservation and total abundance of the radiolarian assemblages examined were also determined for each sample. Preservation is ranked as follows: Poor (P) = over 50 per cent of the specimens are broken and/or show signs of dissolution; moderate (M) = 50 to 25 per cent of the specimens are broken and/or show signs of dissolution; and good (G) = less than 25 per cent of the specimens are broken and/or show signs of dissolution. Abundance is characterized as follows: rare (R) = radiolarians comprise less than 5 per cent of all the sediment material on the slide; few (F) = radiolarians comprise between 5 to 33 per cent of all the sediment material on the slide; and common (C) = radiolarians comprise more than 33 per cent of all the sediment material on the slide. To make these ratings meaningful the following average values were assigned: A slide with moderate preservation and an abundance measurement of few contains approximately 200 radiolarians, with a range of from about 120 to 280.

¹ Present address: Getty Oil Company, Exploration and Production Research Center, Houston, Texas.

TABLE 1
Abundance/Occurrence Chart, DSDP Hole 438

Tables 1 through 8 summarize the foregoing measurements and observations and give the reader an idea of relative abundance and consistency of occurrence of the species used for biostratigraphic analysis. It is from these tables that the list of datums recognized in each hole is constructed (see Table 9).

Table 9 was developed using the maximum range of the selected species and placing the first (B) or last (T) occurrence datum of that species in its proper relative position, except when there was obvious reworking. Transitions from one species to another—for example, from *Stichocorys delmontensis* to *Stichocorys peregrina*—are also reported and represent the stratigraphic level at which the descendant became more abundant upsection, in the same sense as the evolutionary boundaries of Riedel and Sanfilippo (1971). Core barrel and section numbers, with their associated depths in meters below the sediment surface, indicate the location of each datum recognized in the different holes. Depths should increase from the top of the table to the bottom. Arrows indicate if a datum is depressed (↓) or elevated (↑) in the hole relative to the observed maximum range of the species. Elevations of bottoms and depressions of tops are accounted for by poor representation of that particular species in the hole. The number of holes containing arrows for a particular datum indicates the reliability of that datum.

Range charts (Figures 1, 2, and 3) are produced from Tables 1 to 9. Figure 1 is a range chart for Hole 438A. The dashed lines indicate the maximum range of a species as interpreted from the other holes. Figure 2 shows the range of several Pleistocene species found in the upper part of Holes 440 and 440A and its relationship to other radiolarian biozonations. A range chart is produced for the lower portion of Hole 438B for the middle and lower Miocene (see Figure 3). The dashed lines are inferred ranges extrapolated from Holes 438A and 439. These charts are presented to aid in the use of the proposed radiolarian zonation.

BIOSTRATIGRAPHY

The radiolarian zonations proposed by Riedel and Sanfilippo (1970, 1971, 1977, 1978) were not applicable to the samples examined from Leg 57, primarily because of the influx of cold water into the study area by the Oyashio Current from the north. However, several of the datums in their zonations were present and are used to correlate these zonations to the one developed in this study (see Figure 4).

As an alternative to applying the Riedel and Sanfilippo zonations, those developed and/or used by Hays (1970), Kling (1973), and Forman (1975) were implemented. These biozonal schemes extend from upper Miocene to Recent. Older strata had previously been fitted into the zonal scheme of Riedel and Sanfilippo (Kling, 1973; Forman, 1975). These assignments relied on the presence of such warm-water radiolarians as *Cannartus petterssoni* and *Ommatartus antepenultimo*, which appear to invade high-latitude waters. Placement of strata into a radiolarian zone which does not contain the nominate species requires indirect correlation. For example,

Kling (1973) placed the lower portion of DSDP Site 173 in the *Calocyctella costata* Zone, though he lacked the nominate species, *C. costata*, and did not recognize the transition of *Dorcadospiris dentata* to *D. alata*. The presence of *Cannartus violina* indicated that this section might be of the *Calocyctella costata* Zone.

In order to avoid such difficulties, I have presented a new Neogene radiolarian biozonation. Correlation of the radiolarian zonation of Riedel and Sanfilippo (1977, 1978) to the zonation presented herein was furnished using the datums listed in Table 9. This Neogene radiolarian zonation is composed of 16 zones, 9 of which are new. Use of this temperate radiolarian zonation in other high- and low-latitude regions appears to be favorable, because it is based primarily on species present in warm-water as well as in cold-water regions. Therefore it may serve as a link between high- and low-latitude radiolarian zonations.

Description of Zonation

Botryostrobus aquilonaris Zone (Hays, 1970)

The base of the *Botryostrobus aquilonaris* Zone is defined by the last occurrence of *Axoprunum angelinum*, and the top extends into Recent sediments. The *B. aquilonaris* Zone as used herein is equivalent to the *Artostrobium miralestensis* Zone of Kling (1973), the *A. tumidulum* Zone of Forman (1975), and the *Eucyrtidium tumidulum* of Hays (1970). The last occurrence of *Stylacontarium acquilonium* occurs slightly above the base of this zone. An absolute age of 0.4 m.y. has been assigned to the base of the zone by Hays (1970). Sites 438 (Sections 438-1-1-438-2-5 and 438A-1-1-438A-1, CC), 440 (Sections 440-1-1-440A-1-5), and 441 (Sections 441-1-1-441-1, CC) contain strata representing this zone.

Axoprunum angelinum Zone (Kling, 1973)

The top of this zone is defined by the last common occurrence of *Axoprunum angelinum* and its base by the last occurrence of *Eucyrtidium matuyamai*. It is equivalent to the *Axoprunum angelinum* Zones of Kling (1973) and Forman (1975) and the *Stylarctus universus* Zone of Hays (1970). The first occurrence of *Lynchocanium* sp. cf. *L. grande* occurs near its base. This zone is recognized at Sites 438 (Sections 438-3-1-438-5-1 and 438A-2-1-438A-3-5), 440 (Section 440A-3-1-440B-7-3), and 441 (Section 441-2-1). Hays (1970) has correlated the base to the Jarmillo Event (0.9 m.y., Cox, 1969). Hence the zone is assigned to the Pleistocene.

Eucyrtidium matuyamai Zone (Hays, 1970 emend. Kling, 1973)

The base of this zone is defined by the transition of *Eucyrtidium calvertense* to *E. matuyamai*, and the top is equivalent to the base of the *Axoprunum angelinum* Zone. The *Eucyrtidium matuyamai* Zones of Kling (1973) and Forman (1975) are synonymous with it and correlate in part with Hays' (1970) *E. matuyamai* Zone. Sites 438 (Sections 438-5-3 and 438A-4-1-438A-4-5), 440 (Section 440B-7-5-440B-19-3), and 441 (Sections

TABLE 2
Abundance/Occurrence Chart, DSDP Hole 438A

Sample (Interval in cm)	Abundance	Preservation	Collosphera sp. A <i>C. pyrena</i>	<i>Axoprunum angelinum</i> <i>Sphaeropyle langii</i> <i>S. robusta</i>	<i>Stylacantharium acutum</i> <i>S. sp. cf. S. acutum</i> <i>Cenaria laticonus</i> <i>C. mammiferus</i> <i>C. (?) pettersoni</i> <i>C. violina</i> <i>Omnifarctus antipenitimus</i> <i>O. hughesi</i> <i>O. penitimus</i> <i>O. sp.</i>	<i>O. tetratalamus</i> <i>Amphiroplatum ypsilon</i> <i>Spongaster pentas</i> <i>S. tetras tetras</i> <i>Lithocarpium polyacantha</i>	<i>Pranoplite titan</i> <i>P. titan Form A</i> <i>Lamprocyclus maritilis maritilis</i> <i>L. maritilis polypora</i> <i>Lamprocyrts (?) hanai</i>	<i>L. hasti</i> <i>L. heteroporus</i> <i>Pleocorys clausus</i> <i>Thecocorythium trachellum dianae</i> <i>T. vetulum</i>	<i>Aritotrobus annulatus</i> <i>A. (?) pretabulatus</i> <i>Borystrobus aquilonaris</i> <i>B. auritus-australis</i> <i>B. bromleitei</i>	
438A-1-1, 40-42	C C G G	Reworking	1 1	1 2 2 2 2 1						
438A-1, CC	F G G G									
438A-2-1, 26-28	F G G G									
438A-2-3, 6-8	F G G G									
438A-2-5, 30-32	F G G G									
438A-3-1, 124-126	R M M M									
438A-3-3, 35-37	R M M M									
438A-3-5, 30-32	F G G G									
438A-4-1, 77-79	F G G G									
438A-4-3, 50-52	F G G G									
438A-4-5, 40-42	C G G G	*		1 2 1 1	3					
438A-5-1, 40-42	F G G G			2 2	2					
438A-5-2, 68-70	F G G G			2 1 1	2					
438A-5-5, 31-33	F G G G									
438A-6-1, 53-55	F G G G									
438A-6-3, 20-22	F G G G	*	1		1					
438A-6-5, 18-19	F G G G	*	1		1					
438A-7-1, 120-122	F G G G	*	1	1	1					
438A-7-3, 50-52	F G G G									
438A-7-5, 32-34	F G G G									
438A-8-1, 37-39	F G G G	*	1		1					
438A-8-3, 19-21	F G G G	*	1		1					
438A-8-5, 44-46	F G G G	*	1		1					
438A-9-1, 19-21	F G G G	*	1		1					
438A-10-1, 17-19	R G G G									
438A-10-3, 67-69	F G G G	*		1	1					
438A-10-5, 38-40	F G G G	*		1	1					
438A-11-1, 41-43	F G G G	*		1	1					
438A-11-3, 41-43	F G G G	*		1	1					
438A-11-5, 41-43	F G G G	*		1	1					
438A-12-1, 44-46	F G G G	*	1	1	1					
438A-13-1, 27-29	F G G G	*	1	1	1					
438A-13-3, 27-29	F G G G	*	1	1	1					
438A-13-5, 14-16	R G G G	*	1	1	1					
438A-14-1, 34-36	R G G G	*	1	1	1					
438A-14-3, 34-36	C G G G		2	1	1					
438A-14-5, 34-36	C G G G		1	1	1					
438A-15-1, 26-28	C G G G		2	1	1					
438A-16-1, 48-50	F G G G		1	1	1					
438A-16-3, 48-50	F G G G		1	1	1					
438A-16-5, 48-50	C G G G		2	1	1					
438A-18-1, 20-22	F G G G		1							
438A-18-3, 20-22	F G G G		2							
438A-19-1, 20-22	C G G G		1							
438A-19-3, 20-22	C G G G		1							
438A-19-5, 18-20	F G G G		2		1					
438A-20-1, 32-34	C G G G		1		1					
438A-20-3, 32-34	C G G G		1		1					
438A-21-1, 50-52	F G G G		1		1					
438A-21-3, 50-52	C G G G		1		1					
438A-21-5, 30-32	C G G G		1	2	1	1			1	?
438A-22-1, 10-12	C G G G		2	1	1	1		1	1	1
438A-22-3, 10-12	C G G G		2	1	1	1		1	1	1
438A-23-1, 25-27	C G G G		2			1		1	1	1
438A-24-1, 30-34	C G G G		2	1	1	2		1	1	1
438A-24-3, 30-34	C G G G		2			1		1	1	1
438A-24-5, 26-28	C G G G		1	1		1		1	1	1
438A-25-1, 16-18	C G G G		2	1	1	1		1	1	1
438A-25-3, 24-26	C G G G		1	1	1	1		1	1	1
438A-25-5, 24-26	C G G G		1	1	1	2		1	1	1
438A-25-7, 40-42	C G G G		1	2	1	1		1	1	3
438A-26-1, 24-26	C G G G		1	3	1	1		1	1	3
438A-26-3, 30-32	C G G G		1	3	1	1		1	2	3
438A-26-5, 28-30	C G G G		1	1	1	2		1	1	2
438A-27-1, 30-32	F G G G	*	1	1		1		1	1	2
438A-27-3, 30-32	C G G G		1					1	1	3
438A-27-5, 30-32	F G G G		1					1	1	2
438A-28-2, 30-32	F G G G		1					1	1	1
438A-28-4, 30-32	F G G G		1					1	1	2
438A-28-6, 30-32	F G G G		1	1				1	1	1

TABLE 2 - *Continued*

<i>B. miralestensis</i>																				
<i>Phormostichocortus corbulia</i>																				
<i>P. dolulum</i>	1	1																		
<i>P. fistula</i>	1	1																		
<i>Siphocampae nodosaria</i>	1	1																		
<i>Siphositchartus corona</i>																				
<i>Spirocyritis gyrosularis</i>																				
<i>S. scalaris</i>																				
<i>S. subcircularis</i>																				
<i>Clathrocytus bicornis</i>																				
<i>C. cabilloniensis</i>																				
<i>Cycladophora davisiана cornutoides</i>																				
<i>C. davisiана</i>																				
<i>Cyrtocapsella cornuta</i>																				
<i>C. japonica</i>																				
<i>C. tetrapera</i>																				
<i>Eucyrtidium acuminatum</i>																				
<i>E. anomalam</i>																				
<i>E. calverense</i>																				
<i>E. hexagonatum</i>																				
<i>E. inflatum</i>																				
<i>E. matuyamai</i>																				
<i>Lipmanella</i> sp. aff. <i>Thecoctrys redondoensis</i>																				
<i>Lithocanpe</i> sp.																				
<i>L. subligata</i>																				
<i>Lithopera bacca</i>																				
<i>L.. neotera</i>																				
<i>L. renzae</i>																				
<i>Lychnocarium grande</i>																				
<i>L.</i> sp. cf. <i>L. grande</i>																				
<i>Pterocanium praetextum eucolepum</i>																				
<i>P. praetextum praetextum</i>																				
<i>P. trilobatum</i>																				
<i>Dicyophimus</i> sp.																				
<i>D.</i> sp. cf. <i>D. hirundo</i>																				
<i>Stictocorys armata</i>																				
<i>S. dimontensis</i>																				
<i>S. diploconus</i>																				
<i>S. peregrina</i>																				
<i>Thecoctrys bicornis</i>																				
<i>T. japonica</i>																				

TABLE 3
Abundance/Occurrence Chart, DSDP Hole 438B

	Sample (Interval in cm)			Abundance	Preservation	Reworking	<i>Collospheara</i> sp. A	<i>C. pyloma</i>	<i>Axoprunum angelinum</i>	<i>Sphaeropyle robusta</i>	<i>Cannartus laticonus</i>	<i>C. mammiferus</i>	<i>C. violina</i>	<i>Ommatartus antepenultimus</i>	<i>O. hughesi</i>	<i>Lithocarpium polycanthia</i>	<i>Prunopyle titan</i>	<i>Theocorythium trachellum dianae</i>	<i>Artostrobus annulatus</i>	<i>A. (?) pretabulatus</i>	<i>Borystrobus auritus-australis</i>	<i>Phormostichochortus fistula</i>	<i>P. marylandicus</i>	<i>Siphocampe nodosaria</i>	<i>Siphostichanthus corona</i>	<i>Spirocyrts subscalaris</i>	<i>Clathrocytis bicornis</i>	<i>C. cabrilloensis</i>	<i>Cycladophora davisiana cornutooides</i>	<i>Cyrtocapsula cornuta</i>	<i>C. japonica</i>	<i>C. tetrapera</i>	<i>Eucyrtidium calvertense</i>	<i>E. hexagonatum</i>	<i>E. inflatum</i>	<i>Lithocampe subligata</i>	<i>Lithopera bacca</i>	<i>L. neotera</i>	<i>L. renzae</i>	<i>Lychnocanum grande</i>	<i>Stichocorys armata</i>	<i>S. delmontensis</i>	<i>S. diploconus</i>	<i>S. peregrina</i>	<i>Theocalyptra bicornis</i>	<i>T. japonica</i>
438B-2, CC	C	G		2	1	1	2	1																																						
438B-3-3, 28-30	F	G		1																																										
438B-4-1, 94-96	F	G		1	1																																									
438B-4-3, 37-40	C	G		1																																										
438B-4-5, 50-53	F	M																																												
438B-5-1, 118-120	F	M																																												
438B-6-1, 98-100	F	M																																												
438B-6-3, 20-24	F	G																																												
438B-7-1, 130-132	F	G																																												
438B-7-3, 101-104	F	G																																												
438B-8-1, 97-100	F	G		1	1	1																																								
438B-9-1, 6-8	F	M																																												
438B-9-3, 50-53	F	M																																												
438B-10-1, 1-2	F	G		1	1																																									
438B-11-1, 82-83	F	G																																												
438B-12-1, 102-103	F	G																																												
438B-12-3, 21-23	R	M																																												
438B-12-5, 52-54	F	M																																												
438B-13-1, 35-37	F	M		1	1																																									
438B-13, CC	F	M																																												
438B-15-2, 65-66	F	M																																												
438B-16-1, 86-88	F	M																																												
438B-17-1, 81-83	R	P																																												
438B-17-3, 30-33	R	P																																												
438B-18-1, 123-126	R	P																																												
438B-18-3, 73-75	R	P	*																																											
438B-18-5, 41-44	R	P																																												
438B-19-1, 130-133	R	P																																												
438B-19-3, 130-133	R	P																																												
438B-19-5, 125-128	R	P	*																																											
438B-20-1, 86-88	R	P	*																																											
438B-20-3, 97-100	R	P	*																																											
438B-20-5, 14-16	R	P																																												
438B-21-1, 68-70	R	P																																												
438B-21-3, 86-88	R	P																																												
438B-22-1, 49-51	R	P	*																																											
438B-23-1, 74-75	R	P	*																																											
438B-23, CC	R	P	*																																											
438B-24, CC	R	P																																												

TABLE 5
Abundance/Occurrence Chart, DSDP Hole 440

	Sample (Interval in cm)	Abundance	Preservation	Reworking	
440-1-1, 120-122	C G	*	<i>Buccinosphaera invaginata</i>		
440-1-3, 13-15	F G	1	<i>Collosphera</i> sp. A		
440-2-1, 143-145	C G	1	C. sp. B		
440-2-3, 130-132	C G	1	<i>C. tuberosa</i>		
440-2-5, 106-108	C G	1			
440-3-1, 110-112	F G	1			
440-3-3, 93-95	C G	1			
440-4-1, 130-132	C G	1			
440-4-3, 131-133	F G	1			
440-4-5, 120-121	C G	*			
440-5-1, 123-125	C G	1			
440-5-3, 135-140	F G	*			
440-5-5, 34-36	C G	1			
440-6-1, 14-16	C G	*			
440-6-3, 40-42	C G	1			
440-6-5, 28-30	C G	1			
440-7-1, 140-142	C G	1			
440-8-1, 100-102	C G	1			
440-8, CC	C G	1			

TABLE 6
Abundance/Occurrence Chart, DSDP Hole 440A

	Sample (Interval in cm)	Abundance	Preservation		
440A-2-1, 132-134	C G	2	<i>Axoprunum angelinum</i>		
440A-2-3, 130-132	C G	1	<i>Collosphaera</i> sp. A		
440A-2-5, 50-52	F G	1	<i>Sphaeropyle langii</i>		
440A-3-1, 130-132	C G	1	<i>Stylacantharium acquinonium</i>		
440A-3-3, 98-100	C G	1	<i>Omnitaritus tetrahalamus</i>		
440A-4-1, 130-132	C G	1	<i>Amphirhopalum ypsilon</i>		
440A-4, CC	F M	1	<i>Spongaster tetrastriatus irregularis</i>		
440A-5-3, 110-112	C G	1	<i>S. tetras tetras</i>		
440A-5-5, 30-32	C G	1	<i>Lamprocyclas maritilis maritilis</i>		
440A-6-2, 50-52	C G	1	<i>Pterocyclus clausus</i>		
440A-7-1, 44-46	C G	1	<i>Thecothyridium trachellum dianae</i>		
440A-7-3, 7-9	C G	1	<i>Ariostrobus annulatus</i>		
440A-7-5, 49-51	C G	1	<i>Ariostrobus (?) pretabulatus</i>		
440A-7, CC	C G	2	<i>Bostryostrobus equilobularis</i>		
			<i>B. australis-australis</i>		
440A-2-1, 132-134	C G	2	<i>Phormostichoarius fistula</i>		
440A-2-3, 130-132	C G	1	<i>Siphocampe nodosaria</i>		
440A-2-5, 50-52	F G	1	<i>Spirocyritis gyroscalaris</i>		
440A-3-1, 130-132	C G	1	<i>S. scalaris</i>		
440A-3-3, 98-100	C G	1	<i>S. subscalaris</i>		
440A-4-1, 130-132	C G	1	<i>Clathrocyclas bicornis</i>		
440A-4, CC	F M	1	<i>Cycladophora davisiана cornutoidea</i>		
440A-5-3, 110-112	C G	1	<i>C. davisiана davisiана</i>		
440A-5-5, 30-32	C G	1	<i>Eucyrtidium acuminateum</i>		
440A-6-2, 50-52	C G	1	<i>E. anomulum</i>		
440A-7-1, 44-46	C G	1	<i>E. calvertense</i>		
440A-7-3, 7-9	C G	1	<i>E. hexagonatum</i>		
440A-7-5, 49-51	C G	1	<i>E. matuyamai</i>		
440A-7, CC	C G	2	<i>E. calvertense</i>		

441-2-3-441-2, CC) all contain strata representing this zone. The transition from *Sphaeropyle robusta* to *S. langii* postdates but nearly coincides with its base.

Lamprocyrtis heteroporos Zone (Hays, 1970 emend. Kling, 1973)

The base of this zone is defined by the last occurrence of *Stichocorys peregrina* (2.8 m.y., Hays, 1970). Near the top, which is defined as the transition from *Eucyrtidium calvertensis* to *E. matuyamai*, the following important datums may be found: the first appearance of *E. matuyamai* (1.8 m.y., Kling, 1973) and the coeval last

occurrences of *Prunopyle titan*, *Lamprocyrtis heteroporos*, and *Clathrocyclas cabrilloensis*. The *L. heteroporos* Zone of Hays (1970) correlates in part with this zone. The *L. heteroporos* Zones of Kling (1973) and Forman (1975) are equivalent to it. The *L. heteroporos* Zone correlates partly with the lower part of the *Pterocanum prismatum* Zone of Riedel and Sanfilippo (1977). It is Pleistocene at its uppermost level but is predominantly Pliocene in age. Sites 438 (Sections 438-5-438-12-5 and 438A-5-1-438A-12-1), 440 (Sections 440B-19-5-440B-35-1), and 441 (Sections 441-3, CC-441-7-1) possess intervals representative of this zone.

***Sphaeropyle langii* Zone (Forman, 1975)**

The base of this zone is defined as the first occurrence of *Sphaeropyle langii*. The top of this zone and the base of the *Lamprocystis heteroporus* Zone are coincident. Important datums within it are as follows (from top to bottom): The nearly coeval last occurrence of *Stichocorys delmontensis* and *Lithocampe subligata*, last occurrence of *Lithocampe* sp. A, last occurrences of *Theocorys redondoensis* and *Lipmanella* sp. aff. *T. redondoensis*, and coeval first occurrence of *Lamprocystis heteroporus* and transition from *Stylacontarium* sp. cf. *S. acquilonium* to *S. acquilonium*, which is interpreted as the Miocene/Pliocene boundary.

The *Sphaeropyle langii* Zone of Forman (1975) is equivalent to this zone. Sites 438 (Sections 438-12, CC and 438A-13-1-438A-32-4), 440 (Sections 440B-35-3-440B-55-1), and 441 (Sections 441-7, CC-441A-8-1) contain strata typifying it. It is partly equivalent to the *Spongaster pentas* Zone of Riedel and Sanfilippo (1977).

***Theocorys redondoensis* Zone (new zone)**

The base of this zone is defined by the transition of *Stichocorys peregrina* from *S. delmontensis*. The top is the base of the *Sphaeropyle langii* Zone. Intervals from Sites 438 (Sections 438A-33-2-438A-42-1), 440 (Sections 440B-55-3-440B-63-1), and 441 (Sections 441A-8, CC-441B-2, CC) contain sediment representing this zone. The upper Miocene *Stichocorys peregrina* Zone of Forman (1975) is wholly equivalent to it. The *S. peregrina* Zones of Riedel and Sanfilippo (1977) and Kling (1973) are partially equivalent to it. This is not to imply, however, that the boundaries are time-equivalent. The first occurrence of *Lipmanella* sp. aff. *Theocorys redondoensis* occurs within this zone.

***Ommatartus penultimus* Zone
(Riedel and Sanfilippo, 1970)**

The top of this zone is the base of the *Theocorys redondoensis* Zone and the base is defined as the transition from *Ommatartus antepenultimus* to *O. penultimus*. This upper Miocene zone is synonymous with the *O. penultimus* Zones of Riedel and Sanfilippo (1977) and Forman (1975). Auxiliary datums which appear to be nearly coeval with the top of it are the last occurrences of *O. hughesi*, *O. penultimus* and *O.* sp. B. Holes 438A (Sections 438A-42-3-438A-52-6) and 440B (Sections 440B-63-3-440B-71, CC) contain strata representative of this zone.

***Ommatartus antepenultimus* Zone
(Riedel and Sanfilippo, 1977)**

This upper Miocene zone is defined by the range of *Ommatartus antepenultimus* subsequent to its transition from *Cannartus laticonus* and prior to its transition to *O. penultimus*. The *O. antepenultimus* Zone of Riedel and Sanfilippo (1977) and Forman (1975) are its equivalent. Representative sediment is recognized in Hole 438A (Sections 438A-53-2-438A-59-5). Important datums occurring at or near the base of this zone are the first appearances of *Lithocampe* sp. A, *O.* sp. B, and *O. penultimus*.

***Ommatartus hughesi* Zone (new zone)**

The base of this zone is defined as the transition from *Cannartus petterssoni* to *Ommatartus hughesi*, and the top is the base of the *O. antepenultimus* Zone. The latest occurrence of *Collosphaera pyloma* is present in the lower part of it. The zone is not reported by Riedel and Sanfilippo, because at lower latitudes the transition from *Cannartus petterssoni* to *O. hughesi* and *C. laticonus* to *O. antepenultimus* are coeval. Hole 438A contains an interval (Sections 438A-59-5-438-62-1) representing this zone.

***Lithopera bacca* Zone (new zone)**

The base of this zone is defined by the last occurrence of *Eucyrtidium inflatum* and the top is equivalent to the base of the *Ommatartus hughesi* Zone. The *Cannartus petterssoni* Zone of Riedel and Sanfilippo (1977) correlates with most of the zone. The first occurrence of *C. petterssoni* occurs within the lower part, thus placing it in the middle Miocene. The first appearance of *Lithopera bacca* occurs at the base of the zone, as does the last occurrence of *L. renzae* and *C. mammiferus*. Hole 438A (Sections 438A-63-1-438A-65-1) contains intervals which are representative of this zone.

***Eucyrtidium inflatum* Zone (new zone)**

This zone is defined as the range of *Eucyrtidium inflatum*. Datums occurring within this zone are as follows (from top to bottom): First occurrence of *Stichocorys peregrina*, first occurrence of *Lithopera neotera*, coeval first occurrence of *Botryostrobus bramlettei* and last occurrence of *Prunopyle titan* form A, last occurrence of *Cannartus violina*, first occurrence of *P. titan* form A, and first occurrence of *L. renzae*. Strata of Holes 438A (Sections 438A-65-3-438A-78-3) and 438B (Sections 438B-3-3-438B-4-5) are assigned to this zone. The *E. inflatum* zone correlates in part with the *Dorcadospiris alata* Zone of Riedel and Sanfilippo (1977). Therefore it is partly middle Miocene.

***Sphaeropyle robusta* Zone (new zone)**

This zone is defined by the range of *Sphaeropyle robusta* prior to the first occurrence of *Eucyrtidium inflatum*. Several datums are nearly coeval with the base of the zone and may be useful in recognizing the base. They are as follows: First appearance of *Collosphaera* sp. A, *Clathrocyclas cabrilloensis*, *Clathrocyclas bicornis*, *Cannartus mammiferus*, and transition from *C. violina* to *C. mammiferus*. The zone is in part correlative with the *Calocyctella costata* Zone of Riedel and Sanfilippo (1977), indicating that it is at least in part lower Miocene. Intervals in Holes 438A (Sections 438A-79-2-438A-82, CC), 438B (Sections 438B-5-1-438B-10-1) and 439 (Section 439-11-1) are assigned to it.

***Lithocarpium polyacantha* Zone (new zone)**

This zone is defined by the range of *Lithocarpium polyacantha* prior to the first occurrence of *Sphaeropyle robusta*. Two datums are present within it. The uppermost is the first occurrence of *Theocalyptra bicornis* and the other is the first occurrence of *Collosphaera*

TABLE 7
Abundance/Occurrence Chart, DSDP Hole 440B

Sample (Interval in cm)	Abundance	Preservation	Reworking	<i>Collosphaera</i> sp. A	<i>Axopnatum angelinum</i>	<i>Sphaeropyle langii</i>	<i>S. robusta</i>	<i>Stylacantharium acquilonium</i>	<i>S. sp. cf. S. acquilonium</i>	<i>Omnitartus tetrathalamus</i>	<i>Amphirhopalum ypsilon</i>	<i>Spongaster pentas</i>	<i>S. tetras irregularis</i>	<i>S. tetras tetras</i>	<i>Lithocarpium polyacantha</i>	<i>Prunopyle titan</i>	<i>Lamprocyclas maritatis maritatis</i>	<i>Lamprocrytis (?) hanmai</i>	<i>L. haysi</i>	<i>L. heteroporus</i>	<i>L. neoheteroporus</i>	<i>Pterocrys clausus</i>	<i>Thecocorythium trachelium dianae</i>	<i>T. trachelium trachelium</i>	<i>T. vetulum</i>	<i>Artostrobus annulatus</i>	<i>A. (?) pretabulatus</i>	<i>Botryostrobus aquilonaris</i>	
440B-1-1, 100-102	C	G		1						2	1																		
440B-1, CC	C	G		2	4		2			2	1																		
440B-3-1, 30-32	R	G			1					1																			
440B-3-3, 50-52	C	G			2		1			1																			
440B-3-5, 26-28	F	M		1	2			2		2	1																		
440B-4-1, 140-142	C	G	*	1	1	1	1			2	1																		
440B-4-3, 45-47	C	G	*	1	2	2	• 1			2	1	1																	
440B-4-5, 21-24	F	M		1	3		2			1																			
440B-5-1, 49-52	F	M		1	1	1	1			1	1	1																	
440B-5-3, 36-38	C	G	*	1	3	1	1			2	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1		
440B-5-5, 40-42	C	G	*		3					2	1																		
440B-6-1, 68-70	C	G	*	1	3	• 1				1	1																		
440B-6-3, 50-52	C	G		1	3					1																			
440B-6-5, 22-24	C	G		1	2		1			1																			
440B-7-1, 62-64	C	G	*	1	3	• 1				1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1			
440B-7-3, 21-23	C	G		3						1	1																		
440B-7-5, 20-22	C	G	*	2	3	• 1				1	1																		
440B-8-1, 57-58	F	G	*	2	3					1																			
440B-8-3, 100-102	F	G	*	1	3	• 3				1																			
440B-8-5, 81-83	F	G		3	2					1																			
440B-9-1, 27-29	F	G	*	2	2	1				1																			
440B-9-3, 7-9	C	G	*	2	2	• 1				1	1																		
440B-10-1, 145-147	F	M		1	1					1																			
440B-10-3, 33-35	F	M								1																			
440B-10-5, 35-37	C	G	*	1	2	• 1				1																			
440B-10-7, 18-20	C	G	*	3	1	2				1	1																		
440B-11-1, 137-139	C	G	*	1	1	• 2				1	1																		
440B-11-3, 133-135	F	M	*	2	1	3				1																			
440B-11-5, 26-28	F	G								2																			
440B-12-1, 114-116	C	G	*	2	1	• 1				1																			
440B-12-3, 129-131	C	G	*	3	1	1				1																			
440B-12-5, 136-138	C	G	*	3	1	2				1																			
440B-13-1, 95-97	C	G	*	1		• 2				1																			
440B-13-3, 30-32	C	G		2						1																			
440B-14-1, 104-106	F	G		2	1	1				1																			
440B-14-3, 30-32	C	G	*	2	1	1	1			1																			
440B-14-5, 30-32	C	G	*	2	1	1				1																			
440B-15-1, 50-52	C	G	*	1	3	1	1	1		1	1																		
440B-15-3, 130-132	C	G		2	1	1	1			1																			
440B-15-5, 22-24	C	G		3	1	1	1			1																			
440B-15-7, 30-32	F	G	*	2	1					1																			
440B-16-1, 46-48	C	G		3	1	1	1			1																			
440B-16-3, 41-44	C	G		2	1	1	1			1																			
440B-16-5, 20-22	C	G		1	1	1	1			1																			
440B-17-1, 66-68	C	G	*	2	1	1	1			1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1		
440B-17-3, 102-105	C	G		1	2	1				1	1																		
440B-17-5, 22-24	C	G		1		1				1																			
440B-18-1, 58-60	C	G		1		1				1																			
440B-18-3, 30-32	C	G		1	1	1	2			1	1																		
440B-19-1, 61-64	C	G		1	1	1	1			1																			

TABLE 7 - *Continued*

Sample (Interval in cm)	Abundance	Preservation	Reworking	<i>Collophaera</i> sp. A	<i>Axoprumum angelinum</i>	<i>Sphaeropyle langii</i>	<i>S. robusta</i>	<i>Stylocontarium aquilonium</i>	<i>S. sp. cf. S. aquilonium</i>	<i>Ommatarius tetrathalamus</i>	<i>Amphithopatum ypsilon</i>	<i>Spongaster pentas</i>	<i>S. tetras irregularis</i>	<i>S. tetras</i>	<i>Lithocarpium polyacanthia</i>	<i>Prunopyle titan</i>	<i>Lamprocystis (?) hawaii</i>	<i>L. hayesi</i>	<i>L. heteroporus</i>	<i>L. neoheretoporus</i>	<i>Pterocorys clausus</i>	<i>Thecocystium trachelium dianae</i>	<i>T. trachelium trachelium</i>	<i>T. vetulum</i>	<i>Artostrophus annulatus</i>	<i>A. (?) pretabulatus</i>	<i>Bostrychostrobus aquilonaris</i>
440B-19-3, 12-14	C	G		2 1 1 1					1 1	1 1																	
440B-20-1, 60-62	C	G		2 1 1 1					1 1	1 1																	
440B-20-3, 40-42	C	G		2 1 1 1					1 1	1 1																	
440B-20-5, 35-37	C	G		1 1 1 1					1 1	1 1																	
440B-21-1, 36-38	C	G		2 1 1 1					1 1	1 1																	
440B-21-3, 52-54	C	G		1 1 1 1																							
440B-21-5, 44-46	F	M		1 1 1 1																							
440B-22-1, 40-42	C	M		2 1 1 1																							
440B-22-3, 33-35	C	G		2 1 1 1																							
440B-23-1, 59-61	C	G		1 1 1 2																							
440B-23-3, 46-48	C	G		1 1 1 1																							
440B-23-5, 27-29	C	G		1 1 2																							
440B-24-1, 58-60	F	G		1 1 1																							
440B-24-3, 21-23	F	G		1 1 1																							
440B-24-5, 130-132	C	M		1 2					1 1																		
440B-25-1, 20-22	F	G		1 1						1																	
440B-26-1, 46-47	F	M		2 1 1						1																	
440B-27-1, 40-42	C	G	*	1 1 1 1					1 1																		
440B-28-1, 91-93	F	G	*	2 1 1						1 1																	
440B-28-3, 28-30	F	G	*	1 1 1 1						1																	
440B-29-1, 48-50	C	G	*	2 1 1 1						1 1																	
440B-29-3, 81-85	C	G		2 1 1 1						1 1																	
440B-30-1, 23-25	F	M		1 1 1																							
440B-31-1, 117-120	F	M		2 1 1																							
440B-31-3, 20-24	F	M	*	2 1																							
440B-31-5, 100-103	F	M	*	2 1																							
440B-32-1, 39-42	F	G	*	1 1																							
440B-32-3, 7-10	C	G		2 1 1																							
440B-32-5, 21-23	F	G	*	1 1 1																							
440B-33-1, 125-128	C	G	*	2 1 1																							
440B-33-3, 14-16	F	M	*	1 1 2						1																	
440B-34-1, 90-92	F	P	*	1 1																							
440B-34-3, 28-30	F	G		1 1 1 1																							
440B-35-1, 38-40	C	G	*	1 1 2																							
440B-35-3, 20-22	C	G	*	1 1 1 1																							
440B-35-5, 8-10	C	G	*	1 1 1 1																							
440B-36-1, 34-36	C	G		1 1 1																							
440B-36-3, 1-3	F	M		1 1																							
440B-36-5, 4-6	F	G		1 1 1 1																							
440B-37-1, 63-65	C	G		1 1 1 1																							
440B-37-3, 32-34	F	G		1 1 1 1																							
440B-38-1, 70-72	C	G		1 1 1 1																							
440B-38-3, 28-30	F	M		1 1 1 1																							
440B-39-1, 147-149	F	M		1 1 1 1																							
440B-39-3, 88-90	C	G		2 1 2 1					1																		
440B-40-1, 19-21	C	G		1 1 1 1																							
440B-40-3, 9-11	C	G		1 1 1 2																							
440B-40-5, 15-17	C	G		1 1 1 1																							
440B-41-1, 106-108	C	M		1 1 1 1																							
440B-42-1, 116-118	F	M		2 1 1 1																							
440B-42-3, 120-123	F	M		1 1 1 1					1																		
440B-43-1, 60-62	F	M		2 1 1 1																							
440B-43-3, 96-98	F	M	*	2 1 1 1																							
440B-43, CC	C	M	*	2 1 1 1																							
440B-45-1, 38-40	C	P		2 1 1 1																							

TABLE 7 - Continued

	<i>B. auritus-australis</i>																			
1		<i>B. bramlettei</i>																		
1		<i>B. miratesensis</i>																		
2		<i>Phormostichoarts corbula</i>																		
1		<i>P. fistula</i>																		
		<i>Siphocampe arachnea</i>																		
1	2		<i>S. nodosaria</i>																	
1	2		<i>Spirocyritis gyrosclariis</i>																	
1	1		<i>S. scalaris</i>																	
1	2		<i>S. subscalaris</i>																	
			<i>Clathrocyclus bicornis</i>																	
1	2			<i>C. cabrilloensis</i>																
1	1			<i>Cycladophora davisianna cornutoides</i>																
2	3			<i>C. davisianna davisianna</i>																
1	2			<i>Cyrtocapsella cornuta</i>																
1	3			<i>C. japonica</i>																
1	1			<i>Eucyrtidium acuminatum</i>																
1	2			<i>E. anomalum</i>																
1	1			<i>E. calvertense</i>																
2	2			<i>E. hexagonatum</i>																
1	1			<i>E. inflatum</i>																
2	2			<i>E. matuyamai</i>																
1	2			<i>Lithocampus sp.</i>																
1	1			<i>L. sublignata</i>																
1	2			<i>Lithopera bacca</i>																
2	2			<i>Lychnocanium grande</i>																
1	1			<i>I. sp. cf. I. grande</i>																
1	3			<i>Pterocanium praetextum eucolpum</i>																
1	2			<i>P. praetextum praetextum</i>																
1	2			<i>P. trilobum</i>																
				<i>Dictyophimus sp.</i>																
				<i>Stichocorys delmontensis</i>																
				<i>S. peregrina</i>																
				<i>Theocalyptra bicornis</i>																
				<i>Theocorys redondoensis</i>																

TABLE 7 - Continued

Sample (Interval in cm)	Abundance	Preservation	Reworking	<i>Collospelta</i> sp. A	<i>Axoprinum angelinum</i>	<i>Sphaeropyle langii</i>	<i>S. robusta</i>	<i>Sylacontarium aquilonium</i>	<i>S. sp. cf. S. aquilonium</i>	<i>Ommatartus tetralthalamus</i>	<i>Amphirhopalum ypsilon</i>	<i>Spongaster pentas</i>	<i>S. tetras irregularis</i>	<i>Lithocarpium polyacantha</i>	<i>Prunopyle titan</i>	<i>Lamprocyris (?) hannai</i>	<i>L. haysi</i>	<i>L. heteroporus</i>	<i>L. neoheretoporus</i>	<i>Pterocorys clausus</i>	<i>Theacorythium trachelium dianae</i>	<i>T. trachelium trachelium</i>	<i>T. vetulum</i>	<i>Artostrobus annulatus</i>	<i>A. (?) pretabulatus</i>	<i>Botryostrophus aquilonaris</i>			
440B-46-1, 87-90	F	P			2	1	1	1																					
440B-46-3, 56-60	F	P			2	1	1	1																					
440B-47-1, 110-113	F	P			2		1	1																					
440B-48-1, 4-7	F	P	*		2	1	1	1																					
440B-48-3, 142-144	F	P	*		2		1	1																					
440B-48-5, 64-67	F	M			1	2	1	1	1		1							1	1	1								1	
440B-49-1, 66-68	F	M			1	2	1	1	1										1	1								1	
440B-49-3, 111-113	F	M			2	1	1	1	1		1								1								1	1	
440B-49-5, 97-99	C	M			2	1	1	1	1										1								1	1	
440B-50-1, 115-117	F	P			2	1	1	1										1	1								1	1	
440B-51-1, 106-108	F	P			2		1		1										1										
440B-51-3, 16-18	F	P	*		2	1	1	1											1	1								1	
440B-52-1, 18-20	C	P			1		1	1											1	1								1	
440B-52-3, 130-132	C	P			2	1	1	1											1	1							1		
440B-53-1, 131-133	F	P			1	1	1	1											1	1							1		
440B-53-3, 111-114	F	P	*		2	1	1	1											1	1							1		
440B-53-5, 134-136	F	P	*		2	1	1	1											1	1									
440B-54-1, 79-81	F	P			2	1	1	1											1	1									
440B-54-3, 101-103	F	P			2	•	1	1	1										1	1							1		
440B-54-5, 2-4	F	P			2		1	1											1	1							1		
440B-55-1, 82-84	F	P	*		2	•	1	1	1										1										
440B-55-3, 38-40	F	P			2		1	1	1										1	1							1	1	
440B-55-5, 86-88	F	P	*		2		1	1	1										1	1									
440B-56-1, 13-15	F	P	*		1		1	1											1	1									
440B-56-3, 76-78	F	P	*		2		1	1											1	1							1	1	
440B-57-1, 19-20	F	P			2		1	1	1										1	1							1		
440B-57-3, 19-21	F	P			1		1	1	1										1								1	1	
440B-57-5, 4-6	F	P			1		1	•	1										1	1							1		
440B-58-1, 20-21	F	P			2		1	1											1	1							1		
440B-58-3, 87-91	F	P			1		1	1	1										1	1							1		
440B-58-5, 3-5	F	P			1		1	•	1										1	1							1		
440B-59-1, 103-104	C	P			2		1		1										1	1							1		
440B-59-3, 41-42	F	P			1			•	1										1	1								•	
440B-59-5, 44-47	F	P			2		1		1										1	1							1		
440B-60-1, 55-57	F	P			2		1	•	1										1	1									
440B-60-3, 126-128	F	P	*		2		1		1										1	1									
440B-61-1, 66-68	F	P			2		1		1										1	1									
440B-62-1, 35-37	F	P			1	2		1											1	1									
440B-63-1, 50-52	F	P			2		1		1										1	1									
440B-63-3, 33-35	F	P			1	2			1										1	1									
440B-64-1, 131-133	F	P			1	1		1											1								1		
440B-65-1, 85-87	F	P			1	1		1											1	1							1		
440B-66-1, 128-130	F	P			1	2		1											1										
440B-66-3, 149-150	R	P			1	1		1											1										
440B-67-1, 120-122	R	P			2														1	1								1	
440B-67-3, 62-64	F	P			1	1		1											1									1	
440B-68-1, 21-22	F	P			1	2		1											1	1								1	
440B-68-3, 37-39	R	P			1		1		1										1										
440B-69-1, 62-66	R	P			1		1	1											1										
440B-69-3, 92-94	R	P			1		1	1											1										
440B-69, CC	R	P	*		1	2		1											1										
440B-71-1, 142-146	R	P			1		1		1										1	1									
440B-71-3, 26-30	F	P			2		1		1										1	1									
440B-71, CC	F	P			1		2		1										1	1	1								

TABLE 7 - *Continued*

<i>B. auritus-australis</i>																					
1		1																			
1		1																			
1		1																			
1		1																			
1		1																			
<i>B. bramlettei</i>																					
<i>B. miralestensis</i>																					
<i>Phormostictoartus corbula</i>																					
<i>P. fistula</i>																					
<i>Siphocampus arachnea</i>																					
<i>S. nodosaria</i>																					
<i>Spirocyritis gyrosularis</i>																					
<i>S. scalaris</i>																					
<i>S. subscalaris</i>																					
<i>Clathrocytus bicornis</i>																					
<i>C. cabrilloensis</i>																					
<i>Cycladophora davisihana cornutoides</i>																					
<i>C. davisihana davisihana</i>																					
<i>Cyrtocapsella cornuta</i>																					
<i>C. japonica</i>																					
<i>Eucyrtidium acuminatum</i>																					
<i>E. anomulum</i>																					
<i>E. cahertense</i>																					
<i>E. hexagonatum</i>																					
<i>E. inflatum</i>																					
<i>E. matiyamae</i>																					
<i>Lithocampus sp.</i>																					
<i>L. subligata</i>																					
<i>Lithopera bacca</i>																					
<i>Lychnocanium grande</i>																					
<i>L. sp. cf. L. grande</i>																					
<i>Pterocanium praetextum eucolpum</i>																					
<i>P. praetextum praeextum</i>																					
<i>P. trilobum</i>																					
<i>Dictyophimus sp.</i>																					
<i>Stichocorys delmontensis</i>																					
<i>S. peregrina</i>																					
<i>Theocalyptra bicornis</i>																					
<i>Theocalyptra redondoensis</i>																					

TABLE 8
Abundance/Occurrence Chart, Holes at Site 441

Sample (Interval in cm)	Abundance	Preservation	Reworking	<i>Collophaera</i> sp. A	<i>C. tuberosa</i>	<i>Axoprinium angelinum</i>	<i>Sphaeropyyle langii</i>	<i>S. robusta</i>	<i>Stylacontarium acquilonium</i>	<i>Cannartus violina</i>	<i>Ommatarius hughesi</i>	<i>O. tetrahalamus</i>	<i>Amphirhopalum ypsilon</i>	<i>Spongaster pentas</i>	<i>S. tetras irregularis</i>	<i>S. tetras tetras</i>	<i>Lithocarpium polyacantha</i>	<i>Prionyple titan</i>	<i>Lamprocyclas maritatis maritatis</i>	<i>I. maritatis polypora</i>	<i>Lamprocyclis (?) hanuai</i>	<i>L. haysi</i>	<i>L. heteroporus</i>	<i>I. neoheteroporus</i>	<i>Pterocyrys clausus</i>	<i>Thecocystis tracheliatum dianae</i>	<i>T. tracheliatum</i>	<i>T. vetulum</i>				
441-1-2, 50-52	C	G	*		1	•	4	•																								
441-1, CC	C	G	*				4																									
441-2-1, 49-52	C	G	*		2	1	•		1																							
441-2-3, 94-96	C	G	*		2	1		1																								
441-2-5, 60-62	C	G	*		3	1		2	1																							
441-2, CC	C	G	*	1		2	2	1	2	1					1		1	1		1	1	2	1	3	1	•						
441-3, CC	C	G	*			1	1	1	1	1							1	1	1	1	1	1	1	3	1							
441-4, CC	C	G	*				1	1	2		1						1	1	1	1	1	1	1	1	1	1						
441-6, CC	C	G	*		2	1	1	1	1								1	1	1	1	1	1	1	1	1	1						
441-7-1, 76-78	C	G	*		1	1	1										1	1	1	1	1	1	1	1	1	1						
441-7, CC	C	G				1	1	2		1									1	1	1											
441-8-1, 59-61	C	G				1	1	1	1										1	1	1											
441-8, CC	F	G			1	1	1	1											1	1	1											
441-9-1, 56-58	F	G			1	1	1	1											1	1	1											
441-9, CC	F	G			1	1													1	1	1											
441A-2-1, 28-30	C	G	*			1	1	1	1	1									1	1												
441A-2, CC	C	G	*			1	1	1	1	1									1	2	1	1										
441A-5-1, 55-57	B																															
441A-5-3, 77-79	C	G	*	1		1	1	1	1																							
441A-5, CC	F	G	*			1	1																									
441A-6, CC	F	G	*				1	1	1	•									1	1												
441A-8-1, 63-65	F	M	*			2	1	1	1	1	•								1	1												
441A-8, CC	F	M				2				1									1	1												
441A-9, CC	R	M				1													1	1												
441A-10-1, 121-123	R	M	*			1													1	1												
441A-11-1, 83-85	R	M	*				1													1												
441A-11, CC	R	M	*																		1											
441A-12-1, 85-88	R	M																														
441A-13-1, 70-72	B																															
441A-13, CC	R	M					1																									
441A-14-1, 44-46	R	M																														
441A-15-1, 46-48	R	M																														
441A-15, CC	R	M																														
441B-1-1, 10-12	R	G	*	1		1	1	1	1	1										1												
441B-1-3, 20-22	R	M	*				1	1	1	1											1											
441B-1, CC	R	M					1	1	1												1											
441B-2-1, 29-31	R	M	*	1		1															1											
441B-2, CC	R	P							1																							

pyloma. Holes 438A (Sections 438A-84-2-438A-86, CC) and 438B (Sections 438B-11-1-438B-13, CC) contain strata assigned to the zone. It probably correlates with part of the *Calocycletta costata* Zone and possibly with part of the *C. virginis* Zone of Riedel and Sanfilippo (1977) as well.

Stichocorys armata Zone (new zone)

This zone is defined by the range of *Stichocorys delmontensis* prior to the first appearance of *Lithocarpium polyacantha*. The base is synonymous with the *Stichocorys delmontensis* Zone of Riedel and Sanfilippo (1978). Riedel's and Sanfilippo's zone could not be recognized in the samples from DSDP Leg 57 because of

the lack of *Stichocorys wolffii*, whose first occurrence defines the top of it. Sediments from Hole 438B (Sections 438B-15-2-438B-17-1) are placed in the *Stichocorys armata* Zone.

Cannartus violina Zone (new zone)

This zone is defined by the range of *Cannartus violina* prior to the first occurrence of *Stichocorys delmontensis*. This lower Miocene zone may correlate with part of the *Cyrtocapsella tetrapera* Zone of Riedel and Sanfilippo (1978). Hole 438B (Sections 438B-17-3-438B-19-5) possesses radiolarian assemblages representative of the zone.

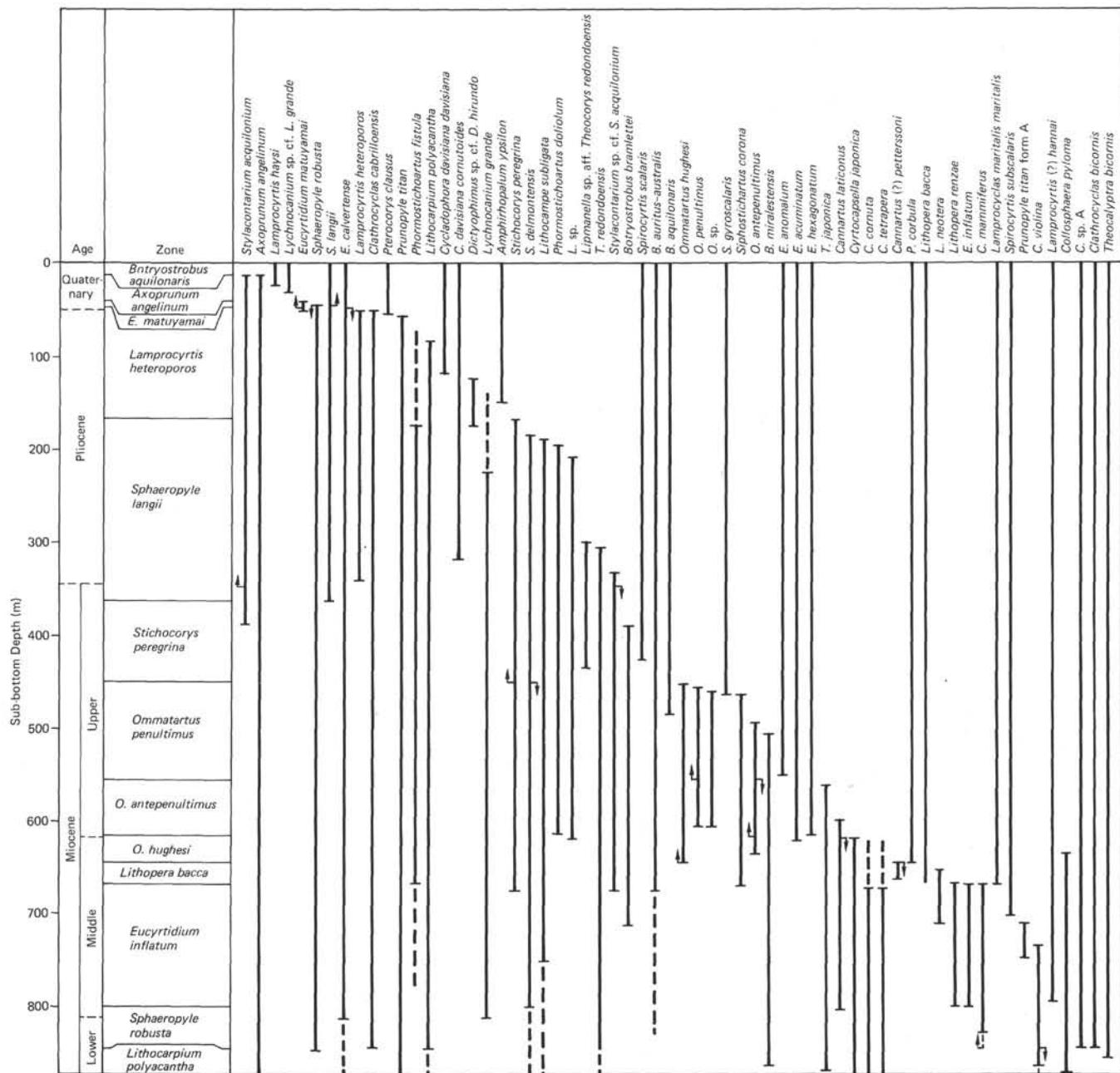


Figure 1. Range chart, DSDP Hole 438A.

Differences in sediment accumulation rates among holes could have been caused by one or a combination of the following factors: (1) differing sedimentation rates, (2) differing amounts of compactions, and (3) the presence of hiatuses.

Reworking

Reworking of older radiolarian assemblages into younger ones proved to be an annoying problem throughout this investigation. This type of sample contamination was usually recognized by the sporadic or isolated occurrence in younger sediments of an extinct species which is continuously present during its normal range — for example, the occurrence of *Eucyrtidium in-*

flatum in lower Pliocene sediments. The problem of determining whether or not sporadically occurring species had been reworked was difficult, because their "true" morphologic tops could not be pinpointed. This may account for the long range of some species.

Tables 1 to 8 list the samples in which reworking is readily recognized. The most intensive reworking was at Site 441, where lower to middle Miocene radiolarians nearly mask the Pleistocene to Pliocene assemblages. Reworking of predominantly Cretaceous and (possibly) early Paleogene radiolarian assemblages into the Oligocene sediments (dated by Keller, this volume) recovered from the lower portion of Hole 439 precluded radiolarian zonal assignments to these strata.

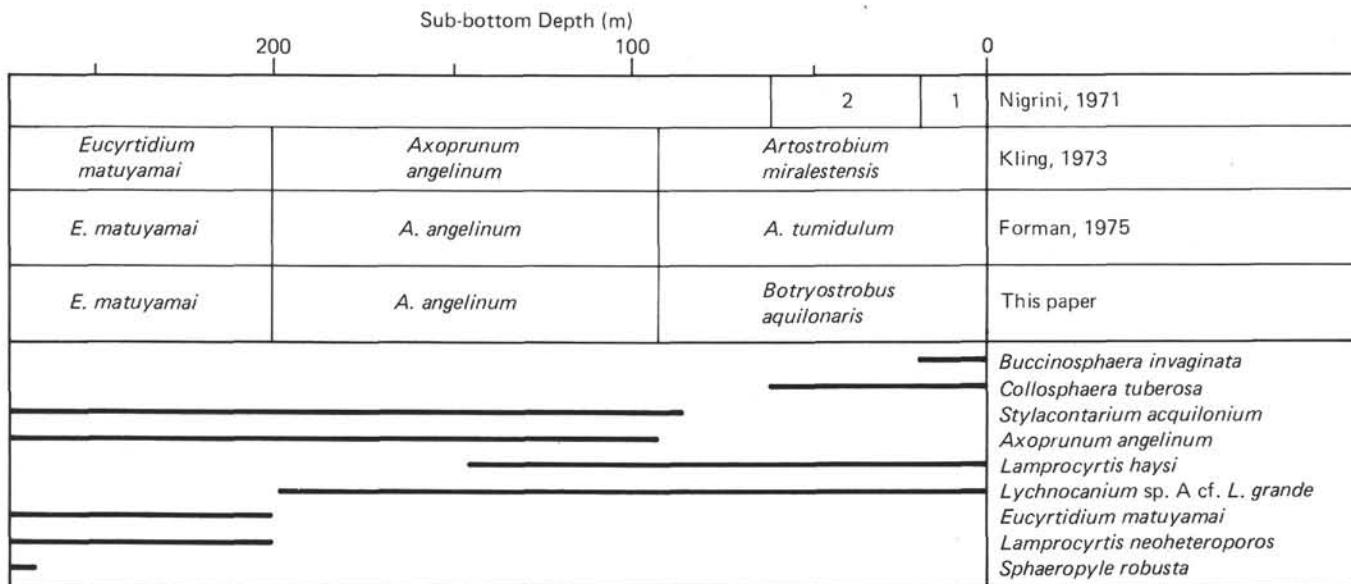


Figure 2. Range chart, DSDP Holes 440 and 440A.

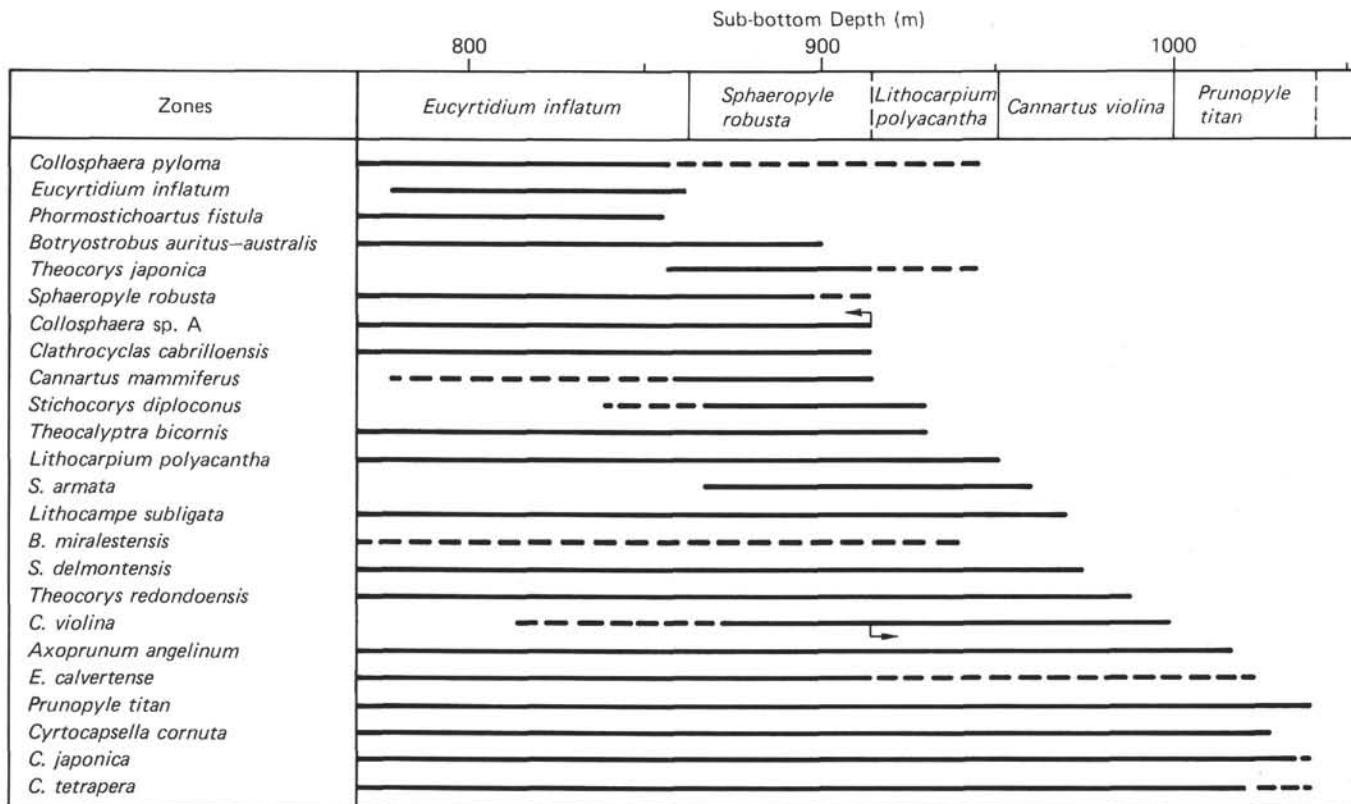


Figure 3. Range chart, DSDP Hole 438B.

Paleotemperatures

Paleotemperature is the only paleocologic factor presented in this report. Paleotemperatures were determined by using the presence or absence of artisiids and selected spongasters (*Spongaster tetras tetras* and *S. pentas*) in a sample. The abundance of these forms collectively was used to determine some minor variations.

Samples were categorized as either warm or cold and no absolute temperatures assigned.

Figure 6 shows the results of this analysis. The asterisks indicate samples in which thin-walled collasphaerids occurred. These forms indicate warm surface waters (Casey, 1972). Their presence within the inferred cold events probably represent pulses of warm surface water into the study area. Figure 6 also shows a paleo-

TABLE 9
Radiolarian Datums

Datums	Hole 438	Hole 438A	Hole 438B	Hole 439	Holes 440, 440A	Hole 440B	Hole 441	Holes 441A, 441B	Zone
B <i>Buccinosphaera invaginata</i>					3-1-3-3 (17-20)				
B <i>Collospira tuberosa</i>					7-1-8-1 (55-65)		1-2-1, CC (2-3)		
T <i>Stylocontarium aquilonium</i>	2-1-2-3 (6-9)	1, CC-2-1 (4-23)			(A) 2-1-2-3 (84-87)		1, CC-2-1 (3-8)		
T <i>Axoprunum angelinum</i>	2-5-3-1 (12-15)	1, CC-2-1 (4-23)			(A) 2-5-3-1 (89-93)		1, CC-2-1 (3-8)		
B <i>Lamprocrytis haysii</i>	3-3-3, CC (18-24)	2-1-2-3 (23-26)				1-1-1, CC (144-149)	1-2-1, CC (2-3)		
B <i>Lychnocanum</i> sp. cf. <i>L. grande</i>	3-3-3, CC (18-24)	2-5-3-1 (29-34)				7-1-7-3 (197-200)	1, CC-2-1 (3-8)		
T <i>Eucyrtidium matuyamai</i>	5-1-5-3 (34-37)	3-5-4-1 (39-43)				7-3-7-5 (200-203)	2-1-2-3 (8-11)		
T <i>Lamprocrytis neoheteroporos</i>						7-3-7-5 (200-203)	2, CC-3, CC (17-26)		
T <i>Sphaeropyle robusta</i>	8-1-8-3 ↑ (63-65)	4-1-4-3 (43-46)				14-1-14-3 (264-266)	2-5-2, CC (14-17)		
<i>S. robusta</i> → <i>S. langii</i>		4-1-4-3 (43-46)				19-1-19-3 (311-314)	2, CC-3, CC (17-26)		
<i>E. calvertense</i>	5-3-5-5 (37-40)	4-3-4-5 (46-49)				19-3-20-1 (314-321)	2, CC-3, CC (17-26)		
→ <i>E. matuyamai</i>	5-3-5-5 (37-40)	4-5-5-1 (49-52)				19-3-20-1 (314-321)	3, CC-4, CC (26-36)		
B <i>E. matuyamai</i>									
T <i>L. heteroporos</i>	5-5-7-1 (40-53)	4-5-5-1 (49-52)				20-3-20-5 (323-326)	2, CC-3, CC ↑ (17-26)		
T <i>Clathrocyclas cabrilloensis</i>	5-5-7-1 (40-53)	4-5-5-1 (49-52)				20-3-20-5 (323-326)			
B <i>Pterocorys clausus</i>	3-3-3, CC ↓ (18-24)	5-1-5-3 (52-55)				12-1-12-3 ↓ (245-248)	3, CC-4, CC (26-36)		
T <i>Prunopyle titan</i>	5-5-7-1 (40-53)	5-3-5-5 (55-58)				14-5-15-1 ↓ (269-273)	2, CC-3, CC ↑ (17-26)		
T <i>Phormostichoartus fistula</i>	7-1-7-3 (53-56)	13-1-13-3 ↑ (173-176)				20-1-20-3 ↑ (321-323)	4, CC-6, CC (36-102)		
T <i>Lithocarpium polycantha</i>	8-1-8-3 (63-65)	5-5-6-1 (58-107)				21-1-21-3 (330-333)	6, CC-7-1 (102-150)		
B <i>Lamprocrytis neoheteroporos</i>						16-5-17-1 ↑ (288-292)	7-1-7, CC (150-159)		
BC <i>Cycladophora davisianna davisianna</i>	9-1-9-3 (72-75)	7-1-7-3 (117-118)				21-1-21-3 (330-333)		(A) 2, CC-5-1 (140-387)	
T <i>Dictyophimus</i> sp.	9-1-9-3 (72-75)	7-5-8-1 (122-126)							
T <i>Lychnocanum grande</i>	10-3-10-5 (85-88)	18-3-19-1 ↑ (221-230)				26-1-27-1 (337-387)	7-1-7, CC (150-159)		
B <i>Amphirhopalum ypsilon</i>	10-3-10-5 (85-88)	10-3-10-5 (148-150)				29-3-30-1 (409-415)	7-1-7, CC (150-159)		
T <i>Stichocorys peregrina</i>	12-5-12, CC (106-109)	12-1-13-1 (164-173)				35-1-35-3 (463-466)	7-1-7, CC (150-159)		
B <i>D.</i> sp.	12-3-12-5 ↓ (104-106)	13-1-13-3 (173-176)							
T <i>S. delmontensis</i>		14-1-14-3 (183-186)				42-3-43-1 ↑ (533-539)			
T <i>Lithocampe subligata</i>		14-5-15-1 (189-192)				36-3-36-5 (475-478)	8-1-8, CC ↑ (207-216)		
T <i>P. doliolum</i>		15-1-16-1 (192-202)							
T <i>L.</i> sp.		16-5-17-1 (208-211)				48-3-48-5 (590-593)	7, CC-8-1 (159-207)		
T <i>Lipmanella</i> sp. aff. <i>Theocorys redondoensis</i>		26-3-26-5 (300-303)							
T <i>T. redondoensis</i>		27-1-27-3 (306-309)		1-1-1, CC ↑ (499-507)		59-5-60-1 ↑ (697-701)			
BC <i>C. davisianna cornutaoides</i>		28-2-28-4 (317-320)				48-1-48-3 ↓ (586-590)		(B) 1, CC-2-1 (334-668)	
T <i>Stylocontarium</i> sp. cf. <i>S. aquilonium</i>		29-6-30-2 (333-336)				54-1-54-3 (644-647)			
B <i>Lamprocrytis heteroporos</i>		30-4-30-6 (339-342)				54-1-54-3 (644-647)		(A) 2-1-2, CC ↓ (130-140)	
S. sp. cf. <i>S. aquilonium</i> → <i>S. aquilonium</i>		31-2-31-4 (347-350)				53-5-54-1 ↓ (638-644)			
B <i>Sphaeropyle langii</i>		32-4-33-2 (359-366)				55-1-55-3 (653-656)			
B <i>Stylocontarium aquilonium</i>		35-4-35-6 (387-390)				57-2-57-5 (675-678)			
T <i>Botryostrobus bramblettii</i>		35-6-36-2 (390-394)				57-5-58-1 (678-681)			
B <i>Spirocrytis scalaris</i>		39-5-40-2 (426-431)				7-5-8-1 ↓ (203-207)	7-1-7, CC ↓ (150-159)		

TABLE 9 – *Continued*

Datums	Hole 438	Hole 438A	Hole 438B	Hole 439	Holes 440, 440A	Hole 440B	Hole 441	Holes 441A, 441B	Zone
B <i>Lipmanella</i> sp. aff. <i>Theocorys redondoensis</i> <i>Stichocorys delmontensis</i> → <i>S. peregrina</i> T <i>Ommatartus hughesi</i> T <i>O. penultimus</i> T <i>Ommatartus</i> sp.		40-4-40-6 (434-437) 42-1-42-3 (449-453) 42-3-42-5 (453-456) 42-5-43-2 (546-460) 43-2-43-4 (460-463)				63-1-63-3 (729-732)			<i>Theocorys redondoensis</i>
B <i>Spirocyrta gyrosularis</i> T <i>Siphostichartus corona</i> B <i>B. aquilonaris</i> T <i>O. antepenultimus</i> T <i>B. miralestensis</i>		43-4-43-6 (463-467) 43-4-43-6 (463-467) 45-3-45-5 (483-486) 46-4-46-6 (493-496) 47-3-47-5 (505-509)				14-3-14-5 (266-269) †		(A) 2, CC-5-1 † (140-387)	<i>Ommatartus penultimus</i>
B <i>E. anomatum</i> <i>O. antepenultimus</i> → <i>O. penultimus</i> T <i>T. japonica</i> B <i>O. penultimus</i> B <i>O.</i> sp.		52-4-52-6 (551-554) 52-6-53-2 (554-557) 53-2-54-2 (557-566) 58-1-59-1 (603-612) 58-1-59-1 (603-612)	4-1-4-3 (854-857) †	1, CC-2-1 (507-556) † 1-1-1, CC (499-507)		58-3-58-5 (685-687) †	9-1-9, CC (264-273) †		
B <i>P. doliolum</i> B <i>E. hexagonatum</i> <i>Cannartus laticonus</i> → <i>O. antepenultimus</i> B <i>E. acuminatum</i> B <i>Lithocampe</i> sp.		59-1-59-3 (612-615) 59-3-59-5 (615-618) 59-5 (618) 59-5-60-1 (618-622) 59-5-60-1 (618-622)		1-1-1, CC (499-507)					<i>Ommatartus antepenultimus</i>
T <i>Cyrtocapsella tetrapera</i> T <i>C. cornuta</i> T <i>C. japonica</i> T <i>Collospheara pyloma</i> B <i>O. antepenultimus</i>		65-3-65-5 † (673-676) 65-3-65-5 † (673-676) 59-5-60-1 (618-622) 60, CC-62-1 (631-641) 60, CC-62-1 (631-641)	6-3-7-1 (875-883) †	2-1-2, CC (556-564) 2-1-2, CC (556-564) 2, CC-3, CC (664-658)					<i>Ommatartus hughesi</i>
T <i>Cannartus petterssoni</i> <i>C. petterssoni</i> → <i>O. hughesi</i> B <i>O. hughesi</i> B <i>P. corbula</i> T <i>Lithopera neotera</i>		62-1-63-1 (641-652) 62-1-63-1 (641-652) 62-1-63-1 (641-652) 62-1-63-1 (641-652) 63-1-64-1 (652-660)	2, CC-3-3 (671-789) 2, CC-3-3 (671-789)	3, CC-4, CC (658-754)					<i>Lithopera bacca</i>
B <i>C. petterssoni</i> B <i>L. bacca</i> T <i>L. renzae</i> B <i>S. corona</i> T <i>E. inflatum</i>		64-3-64-5 (663-666) 64-5-65-1 (666-670) 64-5-65-1 (666-670) 65-1-65-3 (670-673) 65-1-65-3 (670-673)	2, CC-3-3 (671-789)	2-1-2, CC (556-564) †					<i>Lithopera bacca</i>
T <i>C. mammiferus</i> B <i>Lamprocyclas maritalis</i> <i>B. Stichocorys peregrina</i> B <i>Spirocyrta subscalaris</i> B <i>Lithopera neotera</i>		65-1-65-3 (670-673) 65-1-65-3 (670-673) 65-5-66-2 (676-679) 68-1-68-5 (699-705) 68, CC-70-1 (707-717)	4-1-4-3 (854-857) †	2, CC-3, CC (564-658) † 2, CC-3, CC (564-568)					<i>Eucyrtidium inflatum</i>

TABLE 9 – Continued

Datums	Hole 438	Hole 438A	Hole 438B	Hole 439	Holes 440, 440A	Hole 440B	Hole 441	Holes 441A, 441B	Zone
B <i>B. bramlettei</i>		68, CC-70-1 (707-717)		3, CC-4, CC (658-754)					
T <i>Prunoplyte titan</i> form A		68, CC-70-1 (707-717)							
T <i>C. violina</i>		72-1-72-3 (736-739)	5-1-6-1 (864-873) †	10-3-11-1 (900-907) †					
B <i>P. titan</i> form A		73-2-73-4 (747-750)							
T <i>Stichocorys</i> <i>diploconus</i>		75-1-76-1 (764-774)	5-1-6-1 (864-873) †						
B <i>Phormostrichoartus</i> <i>fistula</i>		64-5-65-1 (666-670) †	4-3-4-5 (854-857)						
B <i>Lamprocyrts</i> <i>hannai</i>		78-1-78-3 (794-797)		3, CC-4, CC (658-754)					
B <i>E. inflatum</i>		78-3-79-2 (797-805)	4-5-5-1 (860-864)						
B <i>Lithopera renzae</i>		78-3-79-2 (797-805)							
B <i>C. laticonus</i>		79-2-79-4 (805-808)	2, CC-3-3 (671-879) †	3, CC-4, CC (658-754)					
T <i>S. armata</i>		79-2-79-4 (805-808)	5-1-6-1 (864-873)						
B <i>Lychnocanium</i> <i>grande</i>		79-6-80-2 (811-814)	2, CC-3-3 (671-879) †	3, CC-4, CC (658-754)					
B <i>B. auritus-australis</i>		65-5-66-2 (676-679) †	9-1-9-3 (900-904)	9-3-9-5 (891-894)					
B <i>Sphaeropyle</i> <i>robusta</i>		82, CC-84-2 (840-851)	8-1-9-1 (892-900) †	11-1-11-3 (907-910)					
B <i>Collosphaera</i> sp. A		82, CC-84-2 (840-851)	10-1-11-1 (910-920)	11-1-11-3 (907-910)					
B <i>Clathrocydas</i> <i>cabrilloensis</i>		82, CC-84-2 (840-851)	10-1-11-1 (910-920)	2, CC-3, CC (564-658) †					
B <i>C. bicornis</i>		82, CC-84-2 (840-851)	10-1-11-1 (910-920)	4, CC-5-1 (754-851) †					
<i>Cannartus violina</i> → <i>C. mammiferus</i>		80, CC-82-2 (821-835) †	10-1-11-1 (910-920)						
B <i>Stichocorys</i> <i>diploconus</i>		76-1-77-1 (774-784) †	12-1-12-3 (930-932)						
B <i>Theocalyptra</i> <i>bicornis</i>		84-2-85-2 (851-861)	12-1-12-3 (930-932)	3, CC-4, CC (658-754) †					
B <i>B. miralestensis</i>		85-4-85, CC (864-868)		10-3-11-1 (900-907) †					
B <i>Theocorys</i> <i>japonica</i>		85, CC-86, CC (868-878)	10-1-11-1 (910-920) †						
B <i>Collosphaera</i> <i>pyloma</i>		86, CC (878)	4-3-4-5 (857-860) †						
B <i>Lithocarpium</i> <i>polyacantha</i>		82, CC-84-2 (840-851) †	13, CC-15-2 (948-959)	3, CC-4, CC (658-754) †					
B <i>S. armata</i>		79-4-79-6 (808-811) †	15-2-16-1 (959-967)						
B <i>Lithocampe</i> <i>subligata</i>		73-4-73-6 (750-753) †	16-1-17-1 (967-976)						
B <i>S. delmontensis</i>		78-3-79-2 (797-805) †	17-1-17-3 (976-979)	3, CC-4, CC (658-754) †					
B <i>T. redondoensis</i>		80, CC-82-2 (840-851) †	18-3-18-5 (989-992)	3, CC-4, CC (658-754) †					
B <i>Cannartus violina</i>		85-4-85, CC (864-868) †	19-5-20-1 (1002-1005)	11-3-12-1 (910-917)					
B <i>Axoprunum</i> <i>angelinum</i>									
B <i>E. calvertensis</i>		80-2-80-4 (814-816) †	21-3-22-1 (1018-1024)	13-1-13-3 (926-929)					
B <i>Cyrtocapsella</i> <i>cornuta</i>			10-1-11-1 (910-920)	18-3-19-1 (967-974)					
B <i>C. tetrapera</i>			22-1-23-1 (1024-1034)						
B <i>C. japonica</i>		85, CC-86, CC (868-878) †	23-1-23, CC (1034-1039)	21-3-22-1 (996-1001)					
B <i>P. titan</i>			21-3-22-1 (1018-1024) †	21-3-22-1 (996-1001)					

Eucriodium
*inflatum**Sphaeropyle*
*robusta**Lithocarpium*
*polyacantha**Cannartus*
Stichocorys
*armata**Prunoplyte*
titan

Age	Riedel and Sanfilippo (1977, 1978)	Hays (1970)	Kling (1973)	Forman (1975)	This Paper
Quaternary	no zone	<i>Eucyrtidium tumidulum</i>	<i>Artostrobium miralestensis</i>	<i>Artostrobium tumidulum</i>	<i>Batrystrobus aquilonaris</i>
		<i>Stylactractus universus</i>	<i>Axoprunum angelinum</i>	<i>Axoprunum angelinum</i>	<i>Axoprunum angelinum</i>
		<i>E. matuyamai</i>	<i>Eucyrtidium matuyamai</i>	<i>Eucyrtidium matuyamai</i>	<i>Eucyrtidium matuyamai</i>
Pliocene	<i>Pterocanum prismatum</i>	<i>Lamprocyclas heteroporos</i>	<i>Lamprocyclis heteroporos</i>	<i>Lamprocyclis heteroporos</i>	<i>Lamprocyclis heteroporos</i>
Miocene	<i>Spongaster pentas</i>			<i>Sphaeropyle langii</i>	<i>Sphaeropyle langii</i>
	<i>Stichocorys peregrina</i>			<i>Stichocorys peregrina</i>	<i>Theocorys redondoensis</i>
	<i>Ommatartus penultimus</i>			<i>Ommatartus penultimus</i>	<i>Ommatartus penultimus</i>
	<i>O. antepenultimus</i>			<i>O. antepenultimus</i>	<i>O. antepenultimus</i>
	<i>Cannartus petterssoni</i>			<i>O. hughesi</i>	<i>O. hughesi</i>
	<i>Dorcadospiris alata</i>			<i>Lithopera bacca</i>	<i>Lithopera bacca</i>
	<i>Caiocycletta costata</i>			<i>E. inflatum</i>	<i>E. inflatum</i>
	<i>S. wolffii</i>			<i>S. robusta</i>	<i>S. robusta</i>
	<i>S. delmontensis</i>			<i>Lithocarpium polyacantha</i>	<i>Lithocarpium polyacantha</i>
	<i>Cyrtocapsella tetrapera</i>			<i>Stichocorys armata</i>	<i>Stichocorys armata</i>

Figure 4. Correlation of radiolarian zonations.

temperature curve for Southern California from Casey (personal communication and 1972). A comparison of the two curves reveals that the maximum of the cold (and warm) events are not in phase. This could be due to the poor biostratigraphic control used to correlate the two curves. The phase difference is of the order of 0.25 to 0.5 m.y., with Southern California lagging behind Japan. Therefore the paleotemperatures indicate a cooling (or warming) event first in Japan and then in Southern California. The lag indicates that temperate and/or tropical assemblages are displaced by subarctic and arctic assemblages in Japan first. Moore (1978) shows the expansion of the "Subarctic Factor" (assemblage) and an exclusion of the "Tropical Factor" and "Transitional Factor" during a glacial epoch in the Japan region. For Southern California he shows the "Tropical Factor" and "Transitional Factor" expanding and the "Subarctic Factor" receding during a glacial epoch. Therefore, according to Moore's (1978) data, the two regions may be out of phase by 180° (this aspect is currently being investigated by the author).

TAXONOMIC NOTES

This section provides the reader with a reference containing partial synonymy and/or description and illustration of the species referred to in this study. Remarks clarify the concept of certain species as used herein. The section also provides the description of a new species. The taxonomic framework used closely follows that of Riedel (1971) for the spumellarians and Petrushevskaya (1971) for the nassellarians.

Family COLLOSPHAERIDAE Müller 1858

Buccinosphaera invaginata Haeckel

Buccinosphaera invaginata Haeckel in Nigrini, 1971, p. 445, pl. 34.1, figs. 19, 20.

Collosphaera sp. A (Plate 1, Figures 1-4)

This species possesses the wide platforms between the pores which are typical for the genus, but the wall is typically very robust. The test is always elliptical in shape and is pierced by few to many small pores, which are irregularly disposed. The major and minor axes of this species range between 80 µm to 130 µm and 65 µm to 95 µm respectively. Stratigraphically lower forms tend to be more robust and possess few pores.

Remark: This concept of the species is very general and may in the future be modified to include two species or subspecies. The ranges of these two forms overlap with the stratigraphically lower of the two being more robust than the stratigraphically higher form.

Range: Base of *Sphaeropyle robusta* Zone to Recent.

Collosphaera pyloma n. sp. (Plate 1, Figures 5-9)

This unusual thick-walled collosphaerid possesses a pylome-like structure. The relatively smooth surface of the test is irregularly interrupted by very small circular to subcircular pores of similar size (approximately 3-4 µm). The overall shape of the test is that of a prolate ellipsoid with a minor axis of approximately 80 µm and a major axis of about 110 µm. Small teeth are present at the termination of the pylome.

Range: Lower part of *Lithocarpium polyacantha* Zone to lower part of *Ommatartus hughesi* Zone.

Holotype: Sample 438A-78-3, 90-92 cm, Slide #1, H 14/3 (Plate 1, Figures 7 and 8).

Type locality: Northwest Pacific Ocean, Sample 438A-78-3, 90-92 cm.

Repository: Rice University Micropaleontology Collection.

Collosphaera sp. B

Collosphaera sp. A in Knoll and Johnson, 1975, pl. 1, figs. 1, 2; pl. 2, figs. 4-6, p. 63.

Collosphaera tuberosa Haeckel

Collosphaera tuberosa Haeckel in Nigrini, 1971, p. 445, pl. 34.1, fig. 1.

Family ACTINOMMIDAE Haeckel 1862, emend. Riedel 1967

Axoprunum angelinum (Campbell and Clark)

Axoprunum angelinum (Campbell and Clark) in Kling, 1973, p. 634, pl. 1, figs. 13-16; pl. 6, figs. 14-18.

Sphaeropyle langii Dreyer

Sphaeropyle langii Dreyer in Forman, 1975, p. 618, pl. 9, figs. 30-31.

Remark: *S. langii* is distinguished from *S. robusta* on the basis set forth by Forman, 1975. The second-outermost shell contains large, irregularly arranged pores, fewer than 12 in number on half the circumference.

Sphaeropyle robusta Kling

Sphaeropyle robusta Kling emend. in Forman, 1975, p. 618, pl. 9, figs. 24-26.

Remark: *S. robusta* is distinguished from *S. langii* by having small, regularly disposed pores on the second-outermost shell.

Stylacontarium acquinonum (Hays)

Stylacontarium acquinonum (Hays) in Kling, 1973, p. 634, pl. 1, figs. 17-20; pl. 14, figs. 1-4.

Remark: Individuals categorized under this species contain the protrusions of the elliptical medullary shell along the connecting bars noted by Kling (1973).

Stylacontarium sp. cf. S. acquinonum

Stylacontarium sp. aff. *S. bispiculum* of Kling, 1973, p. 634, pl. 6, figs. 19-23; pl. 14, figs. 5-8.

Remark: Similar to *S. acquinonum* but lacks the protrusions of the elliptical medullary wall along the connecting bars. Kling (1973) noted

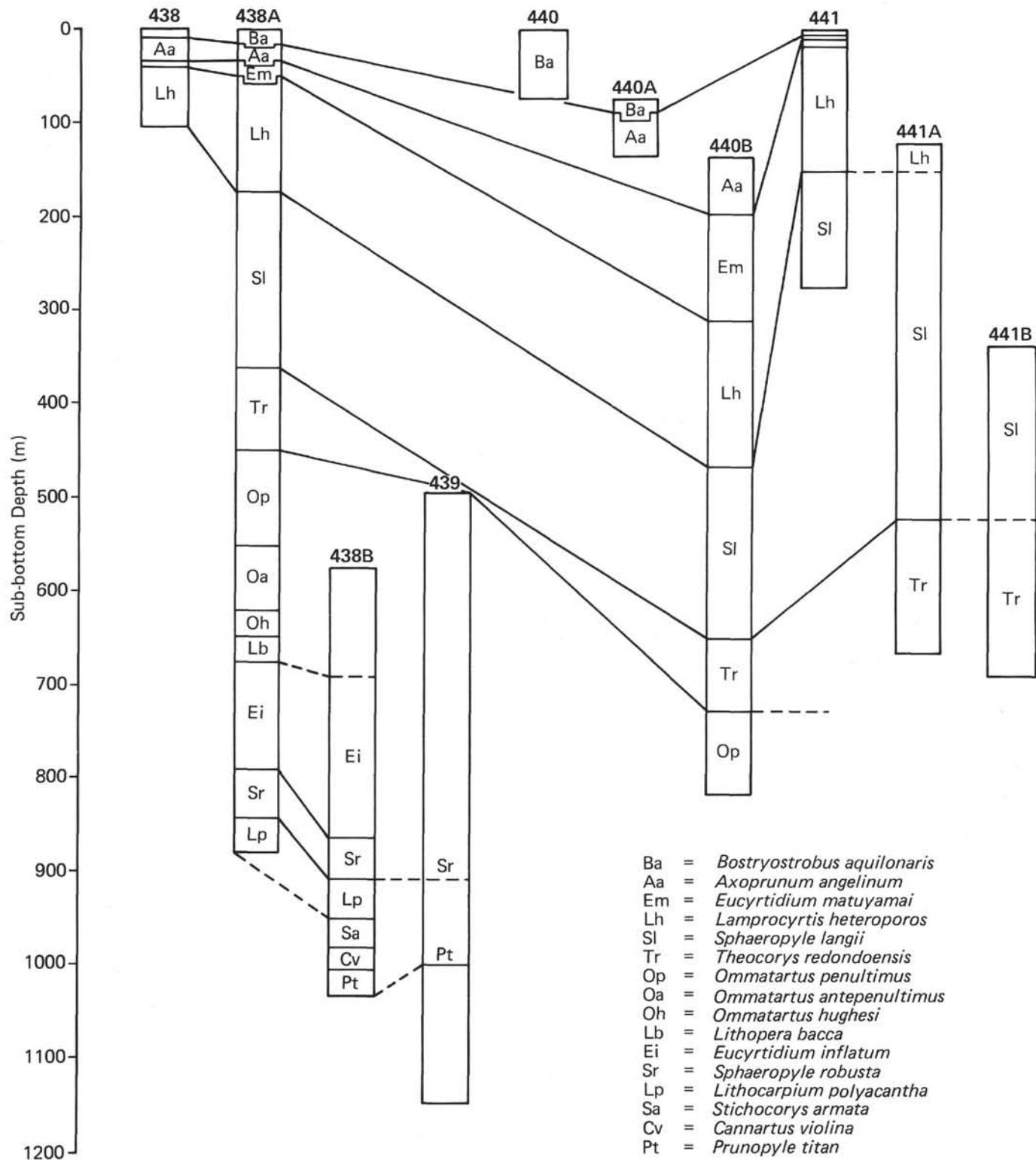


Figure 5. Correlation of holes drilled on DSDP Leg 57.

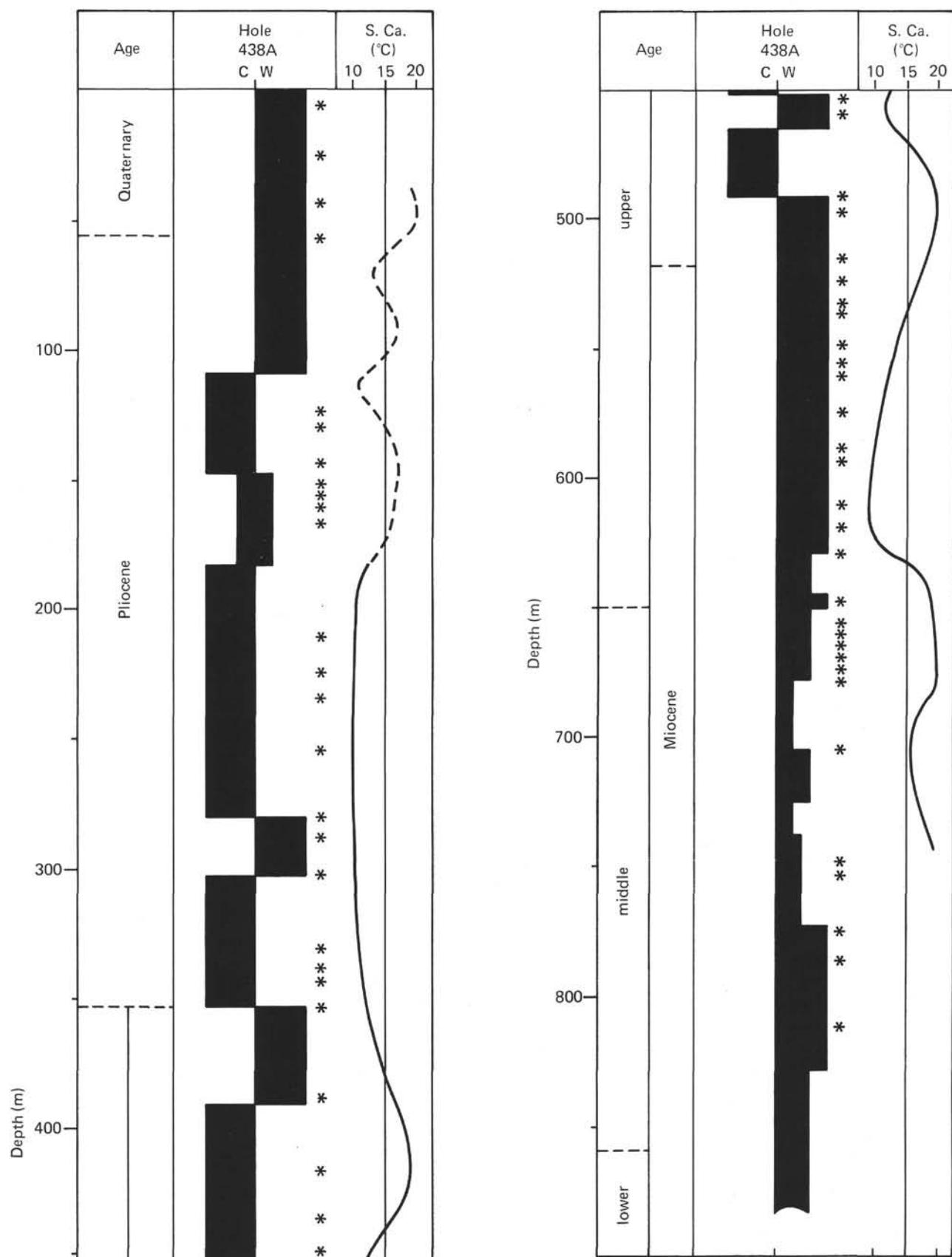


Figure 6. Paleotemperature curves for DSDP Hole 438A and S. California (after Casey, personal communication, and 1972) related to depth in DSDP Hole 438A and its associated age assignments. Asterisks indicate samples containing warm-water collosphaerids.

that this species may be the ancestor of *S. acqilonium*. In this study it is treated as such, and the transition between the two is very close to or at the Miocene/Pliocene boundary.

Subfamily ARTISCINAE Haeckel 1881, emend. Riedel 1967

Cannartus laticonus Riedel

Cannartus laticonus Riedel in Westberg and Riedel, 1978, p. 20, pl. 2, figs. 1-3.

Cannartus mammiferus (Haeckel)

Cannartus mammiferus (Haeckel) in Riedel and Sanfilippo, 1971, p. 1587, pl. 2C, figs. 1-3.

Cannartus (?) petterssoni Riedel and Sanfilippo

Cannartus (?) petterssoni Riedel and Sanfilippo, in Riedel and Sanfilippo, 1971, p. 1587, pl. 1C, figs. 19, 20.

Cannartus violina Haeckel

Cannartus violina Haeckel in Riedel and Sanfilippo, 1971, p. 1588, pl. 2C, figs. 4-7.

Ommatartus antepenultimus Riedel and Sanfilippo

Ommatartus antepenultimus Riedel and Sanfilippo in Westberg and Riedel, 1978, p. 22, pl. 2, figs. 4, 5.

Ommatartus hughesi (Campbell and Clark)

Ommatartus hughesi (Campbell and Clark) in Riedel and Sanfilippo, 1971, p. 1588, pl. 1C, figs. 17, 18.

Ommatartus penultimus (Riedel)

Ommatartus penultimus (Riedel) in Westberg and Riedel, 1978, p. 22, pl. 2, figs. 6-8.

Ommatartus sp.

This species lacks the well-developed equatorial constriction typical of most species of *Ommatartus*. Although some individuals possess small polar caps, most lack any sign of one. May be closely related to *Ommatartus antepenultimus*.

Ommatartus tetrathalamus (Haeckel)

Ommatartus tetrathalamus (Haeckel) in Riedel and Sanfilippo, 1971, p. 1588, pl. 1C, figs. 6, 7.

Family SPONGODISCIDAE Haeckel 1862, emend. Riedel 1967

Amphirhopalum ypsilon Haeckel

Amphirhopalum ypsilon Haeckel in Nigrini, 1971, p. 447, pl. 34.1, figs. 7a-c.

Spongaster pentas Riedel and Sanfilippo

Spongaster pentas Riedel and Sanfilippo, 1970, p. 523, pl. 15, fig. 3.

Spongaster tetras Ehrenberg *irregularis* Nigrini

Spongaster tetras irregularis Nigrini, 1967, p. 43, pl. 5, fig. 2.

Spongaster tetras Ehrenberg

Spongaster tetras Riedel and Sanfilippo, 1971, p. 1589, pl. 1D, figs. 2-4.

Family LITHELIIDAE Haeckel 1862

Lithocarpium polyacantha (Campbell and Clark)

Larnacantha polyacantha Campbell and Clark, 1944 (in part), p. 30, pl. 5, figs. 6, 7.

Lithocarpium polyacantha (Campbell and Clark) group in Petrushevskaya, 1975 (in part), p. 572, pl. 3, figs. 7, 8.

Remark: Presence of inner medullary structures in individuals of this species distinctly distinguish them from individuals of *Prunopyle titan*. Thick cortical walls are common and the surface may have a spinose texture.

Prunopyle titan Campbell and Clark

Prunopyle titan Campbell and Clark, 1944, p. 20, pl. 3, figs. 1-3.

Remark: Included within this species are individuals that lack inner medullary structures. Those containing such structures are placed under *Lithocarpium polyacantha*.

Prunopyle titan Campbell and Clark form A

(Plate 1, Figure 10)

Differs from *P. titan* by its enormous size (860 μm) and more spherical shape. Only a few specimens were found, but it seems useful to separate this form out because of its limited range in the *Eucyrtidium inflatum* Zone.

Family PTEROCORYIDAE Haeckel 1881, emend. Riedel 1967

Lamprocyclas maritatis maritatis Haeckel

Lamprocyclas maritatis maritatis Haeckel in Nigrini, 1967, p. 74, pl. 7, fig. 5.

Lamprocyclas maritatis Haeckel polypora Nigrini

Lamprocyclas maritatis polypora Nigrini, 1967, p. 76, pl. 7, fig. 6.

Lamprocyrts (?) hannai (Campbell and Clark)

Lamprocyrts (?) hannai (Campbell and Clark) in Kling, 1973, p. 638, pl. 5, figs. 12-14; pl. 12, figs. 10-14.

Lamprocyrts haysi Kling

Lamprocyrts haysi Kling, 1973, p. 639, pl. 15, figs. 1-3.

Lamprocyrts heteroporus (Hays)

Lamprocyrts heteroporus (Hays) in Kling, 1973, p. 639, pl. 5, figs. 19, 20.

Lamprocyrts neoheteroporus Kling

Lamprocyrts neoheteroporus Kling, 1973, p. 639, pl. 5, figs. 17, 18; pl. 15, figs. 4, 5.

Remark: This species is not continuously present or abundant in any of the sections examined.

Pterocorys clausus (Popofsky) group

Pterocorys clausus group (Popofsky) in Petrushevskaya and Kozlova, 1972, p. 545, pl. 36, figs. 16-18.

Remark: Individuals of this species have abdomens that are variable (strongly convex to straight). Mouth may be closed or open but is usually constricted.

Theocorythium trachelium (Ehrenberg) *dianae* (Haeckel)

Theocorythium trachelium (Ehrenberg) *dianae* (Haeckel) in Nigrini, 1967, p. 77, pl. 8, figs. 1a, 1b; pl. 9, figs. 1a, 1b.

Theocorythium trachelium trachelium (Ehrenberg)

Theocorythium trachelium trachelium (Ehrenberg) in Nigrini, 1967, p. 79, pl. 8, fig. 2.

Theocorythium vetulum Nigrini

Theocorythium vetulum Nigrini, 1971, p. 447, pl. 1, figs. 6a, 6b.

Family ARTOSTROBIIDAE Riedel 1967

Artostrobus annulatus (Bailey)

Artostrobus annulatus (Baily) in Petrushevskaya, 1975, pl. 10, figs. 4, 5.

Artostrobus (?) pretabulatus Petrushevskaya

Artostrobus (?) pretabulatus Petrushevskaya, 1975, p. 580, pl. 10, figs. 2, 3.

Botryostrobus aquilonaris (Bailey)

Botryostrobus aquilonaris (Bailey) in Nigrini, 1977, p. 246, pl. 1, fig. 1.

Botryostrobus auritus-australis (Ehrenberg) group

Botryostrobus auritus-australis (Ehrenberg) group in Nigrini, 1977, p. 246, pl. 1, figs. 2–5.

Botryostrobus bramlettei (Campbell and Clark)

Botryostrobus bramlettei (Campbell and Clark) in Nigrini, 1977, p. 248, pl. 1, figs. 7, 8.

Botryostrobus miralestensis (Campbell and Clark)

Botryostrobus miralestensis (Campbell and Clark) in Nigrini, 1977, p. 249, pl. 1, fig. 9.

Phormostichoartus corbula (Harting)

Phormostichoartus corbula (Harting) in Nigrini, 1977, p. 252, pl. 1, fig. 10.

Phormostichoartus dololum (Riedel and Sanfilippo)

Phormostichoartus dololum (Riedel and Sanfilippo) in Nigrini, 1977, p. 252, pl. 1, fig. 14.

Phormostichoartus fistula Nigrini

Phormostichoartus fistula Nigrini, 1977, p. 253, pl. 1, figs. 11–13.

Phormostichoartus marylandicus (Martin)

Phormostichoartus marylandicus (Martin) in Nigrini, 1977, p. 253, pl. 2, figs. 1–3.

Siphocampe arachnea (Ehrenberg) group

Siphocampe arachnea (Ehrenberg) group in Nigrini, 1977, p. 255, pl. 3, figs. 7, 8.

Siphocampe nodosaria (Haeckel)

Siphocampe nodosaria (Haeckel) in Nigrini, 1977, p. 256, pl. 3, fig. 11.

Siphostichartus corona (Haeckel)

Siphostichartus corona (Haeckel) in Nigrini, 1977, p. 257, pl. 2, figs. 5–7.

Spirocyrtsis gyrosclaris Nigrini

Spirocyrtsis gyrosclaris Nigrini, 1977, p. 258, pl. 2, figs. 10, 11.

Spirocyrtsis scalaris Haeckel

Spirocyrtsis scalaris Haeckel in Nigrini, 1977, p. 259, pl. 2, figs. 12, 13.

Spirocyrtsis subscalaris Nigrini

Spirocyrtsis subscalaris Nigrini, 1977, p. 259, pl. 3, figs. 1, 2.

Family EUKYRTIDIIDAE Ehrenberg 1847, emend. Petrushevskaya 1971

Clathrocyclas bicornis Hays

Clathrocyclas bicornis Hays, 1965, p. 179, pl. III, fig. 3.

Remark: *Clathrocyclas bicornis* differs from *Theocalyptra bicornis* by having a smaller length-to-width ratio, a much thicker wall, and a different type of internal structure.

**Clathrocyclas cabrilloensis Campbell and Clark
(Plate 1, Figures 15 and 16)**

Clathrocyclas (*Clathrocycloma*) *cabrilloensis* Campbell and Clark, 1944, p. 48, pl. 7, figs. 1, 2.

Remark: Many different forms are grouped under this species, but all of them have surface spines.

Cycladophora davisiана (Ehrenberg) cornutoides Petrushevskaya

Cycladophora davisiана (Ehrenberg) var. *cornutoides* Petrushevskaya, 1967, p. 126, fig. 70, I–III.

Cycladophora davisiана davisiана (Ehrenberg)

Cycladophora davisiана (Ehrenberg) in Petrushevskaya, 1967, p. 122, fig. 69, I–VII.

Cyrtocapsella cornuta Haeckel

Cyrtocapsella cornuta Haeckel in Sanfilippo and Riedel, 1970, p. 453, pl. 1, figs. 19, 20; Kling, 1973, pl. 11, figs. 16–18.

Remark: Individuals without truly hemispherical fourth segments are included in this species. The campanulate shape of the third segment was the principle character used to separate this species from *C. tetrapera*, which has a third segment with nearly parallel walls.

Cyrtocapsella japonica (Nakaseko)

Cyrtocapsella japonica (Nakaseko) in Sanfilippo and Riedel, 1970, p. 542, pl. 1, figs. 13–15.

Cyrtocapsella tetrapera Haeckel

Cyrtocapsella tetrapera Haeckel in Sanfilippo and Riedel, 1970, p. 543, pl. 1, figs. 16–18.

Eucyrtidium acuminatum (Ehrenberg)

Eucyrtidium acuminatum (Ehrenberg) in Kling, 1973, pl. 4, figs. 20–23.

Eucyrtidium anomalum Haeckel

Eucyrtidium anomalum Haeckel, 1862, p. 323, pl. 4, figs. 11–13.

Eucyrtidium calvertense Martin

Eucyrtidium calvertense Martin in Kling, 1973, pl. 4, figs. 16, 18, 19; pl. 11, figs. 1–5.

Remark: Differences in size between *E. calvertense* and *E. matuyamai* was significant and was used to distinguish the two species.

Eucyrtidium hexagonatum Haeckel

Eucyrtidium hexagonatum Haeckel in Nigrini, 1967, p. 83, pl. 8, figs. 4a–b.

Eucyrtidium inflatum

Eucyrtidium inflatum Kling, 1973, p. 535, pl. 11, fig. 7; pl. 15, figs. 7–10.

Eucyrtidium matuyamai Hays

Eucyrtidium matuyamai Hays, 1970, p. 213, pl. 1, figs. 7–9; Kling, 1973, pl. 4, fig. 17.

Lipmanella sp. aff. Theocorys redondoensis

(Plate 1, Figures 17 and 18)

Cephalis is similar to that of *Theocorys redondoensis*. The primary difference between this species and its probable ancestor, *T. redondoensis*, is the lack of a truly spherical and robust thorax and the presence of wings that are initially three-bladed on the thorax of *Lipmanella* sp. aff. *Theocorys redondoensis*.

Lithocampe sp.

(Plate 1, Figure 19)

A five-segmented species with the first four segments similar to those of species of *Cyrtocapsella* and *Stichocorys*. The fifth segment is narrower than the fourth and may be either open or closed. It may be inverted cap-shape or campanulate. This species differs from *Cyrtocapsella tetrapera* and *C. cornuta* in that their fourth segment is closed and from species of *Stichocorys* by the lack of more than five chambers.

Lithocampe subligata Stohr

Lithocampe (*Lithocampe*) *subligata* Stohr group in Petrushevskaya, 1975, p. 581, pl. 14, figs. 6–9, 12.

Lithopera bacca Ehrenberg

Lithopera bacca Ehrenberg in Nigrini, 1967, p. 54, pl. 6, fig. 2.

Lithopera neotera Sanfilippo and Riedel

Lithopera (Lithopera) neotera Sanfilippo and Riedel, 1970, p. 454, pl. 1, figs. 24-26, 28.

Lithopera renzae Sanfilippo and Riedel

Lithopera (Lithopera) renzae Sanfilippo and Riedel, 1970, p. 454, figs. 21-23, 27.

Lychnocanum grande Campbell and Clark

Lychnocanoma grande (Campbell and Clark) in Kling, 1973, pl. 10, figs. 10-14.

Lychnocanum sp. cf. L. grande Campbell and Clark
(Plate 1, Figures 21 and 22)

Lychnocanoma sp. A cf. *L. grande* (Campbell and Clark) in Kling, 1973, pl. 4, figs. 9, 10.

Remark: Individuals placed under this species are quite similar to *L. grande* but usually have a more robust thorax, a campanulate thorax, and smaller pores on the thorax. The abdominal segment seems to be better developed in most individuals than those of *L. grande* and may serve as a criterion in distinguishing the two species. There is an obvious gap in time between the first occurrence of this form and the last occurrence of *L. grande*.

Pterocanium praetextum (Ehrenberg) eucolpum Haeckel

Pterocanium praetextum (Ehrenberg) *eucolpum* Haeckel in Nigrini, 1967, p. 70, pl. 7, fig. 2.

Pterocanium praetextum praetextum (Ehrenberg)

Pterocanium praetextum praetextum (Ehrenberg) in Nigrini, 1967, p. 68, pl. 7, fig. 1.

Pterocanium trilobum (Haeckel)

Pterocanium trilobum (Haeckel) in Nigrini, 1967, p. 71, pl. 7, figs. 3a-b.

Dictyophimus sp.

(Plate 1, Figures 23 and 24)

A dictyophimid with the legs forming from the cephalis and thorax. This form is extremely robust.

Dictyophimus sp. cf. D. hirundo Haeckel

(Plate 1, Figure 20)

Similar to *D. hirundo* except that the thorax is straighter.

Stichocorys armata (Haeckel)

Stichocorys armata (Haeckel) in Riedel and Sanfilippo, 1971, p. 1595, pl. 2E, figs. 13-15.

Stichocorys delmontensis (Campbell and Clark)

Stichocorys delmontensis (Campbell and Clark) in Westberg and Riedel, 1978, p. 22, pl. 3, figs. 1-5.

Stichocorys diploconus (Haeckel)

Stichocorys diploconus (Haeckel) in Sanfilippo and Riedel, 1970, p. 451, pl. 1, figs. 31, 32.

Stichocorys peregrina (Riedel)

Stichocorys peregrina (Riedel) in Westberg and Riedel, 1978, p. 22, pl. 3, figs. 6-9.

Theocalyptra bicornis (Popofsky)

Theocalyptra bicornis (Popofsky) in Riedel, 1958, p. 240, pl. 4, fig. 4.

Theocorys redondoensis (Campbell and Clark)

Theocyrtis (Theocorusca) redondoensis Campbell and Clark, 1944, p. 49, pl. 7, fig. 4.

Theocorys redondoensis (Campbell and Clark) in Kling, 1973, p. 638, pl. 11, figs. 26-28.

Theocorys japonica (Nakaseko)

Sethocyrtis japonica Nakaseko, 1963, p. 176, text fig. 9, pl. 1, figs. 10a, 10b.

Remark: This species differs from *T. redondoensis* in being much smaller in size and in the shape of the thorax. It is probably the ancestor of *T. redondoensis*.

ACKNOWLEDGMENTS

Acknowledgment is made to the Donors of the Petroleum Research Fund, administered by the American Chemical Society, for partial support of this research through PRF Grant 8657-AC2 and to the Oceanography Section of the National Science Foundation for partial support through NSF Grant OCE74-21805. I thank F. M. Weaver for his review of the manuscript and R. E. Casey for his review and continued support in the preparation of the manuscript. C. J. Reynolds is most graciously acknowledged for her typing of the manuscript.

REFERENCES

- Campbell, A. S., and Clark, B. L., 1944. Miocene radiolarian faunas from Southern California. *Geol. Soc. Am. Spec. Paper*, 51, 76.
- Casey, R. E., 1972. Neogene radiolarian biostratigraphy and paleotemperatures: Southern California, the Experimental Mohole, Antarctic core E 14-8. *Palaeogeogr., Palaeoclimatol., Palaeoecol.*, 12, 115-130.
- Cox, A., 1969. Geomagnetic reversals. *Science*, 163, 237-245.
- Ehrenberg, C. G., 1847. Ueber die Mikroskopischen Kisselschaugen Polycystinen als Machtige Gebirgsmasse von Barbados. *K. Preuss. Akad. Wiss. Monatsber.* (for 1847), pp. 40-60.
- Foreman, H. P., 1975. Radiolarians from the North Pacific, Deep Sea Drilling Project, Leg 32. In Larson, R. L., Moberly, R., et al., *Init. Repts. DSDP*, 32: Washington (U.S. Govt. Printing Office), 579-676.
- Haeckel, E., 1862. Die radiolarien (*Rhizopoda Radiolaria*). *Eine monographie*: Berlin (Reimer), p. 572.
- , 1881. Entwurf eines Radiolarien. Systems auf Grund von Studien der Challenger-Radiolarien. *Jenaische Zeitschr. Naturwiss.*, 15, 418-472.
- Hays, J. D., 1965. Radiolaria and late Tertiary and Quaternary history of Antarctic seas. *Biology of Antarctic Seas II: Am. Geophys. Union, Antarctic Res. Series*, 5, 125-184.
- , 1970. The stratigraphy and evolutionary trends of Radiolaria in North Pacific deep-sea sediments. *Geol. Soc. Am., Mem.* 126, pp. 185-218.
- Kling, S. A., 1973. Radiolaria from the eastern North Pacific, Deep Sea Drilling Project, Leg 18. In Kulm, L. D., von Huene, R., et al., *Init. Repts. DSDP*, 18: Washington (U.S. Govt. Printing Office), 617-671.
- Knoll, A. H., and Johnson, D. A., 1975. Late Pleistocene evolution of the collosphaerid radiolarian *Buccinosphaera invaginata* Haeckel. *Micropaleontology*, 21 (No. 1), 60-68.
- Moore, T. C., 1978. The distribution of radiolarian assemblages in the modern and ice-age Pacific. *Mar. Micropaleont.*, 3, 229-266.
- Müller, J., 1858. Über die thalassicollen, Polycystinen und Acanthometren des Mittelmeeres. *Abh. Kgl. Akad. Wiss. Berlin*, pp. 1-62.
- Nakaseko, K., 1963. Neogene Cyrtoidaea (Radiolaria) from the Isozaki Formation in Ibaraki Prefecture, Japan. *Osaka Univ. Sci. Rept.*, 12 (2), 165-198.
- Nigrini, C. A., 1967. Radiolaria in pelagic sediments from the Indian and Atlantic Oceans. *Scripps Inst. Oceanogr. Bull.*, 11, 1-106.

- _____, 1971. Radiolarian zones in the Quaternary of the equatorial Pacific Ocean. In Funnell, B. M., and Riedel, W. R. (Eds.), *Micropalaeontology of the Oceans*: Cambridge (Cambridge University Press), pp. 443-461.
- _____, 1977. Tropical Cenozoic Artostrobiidae (Radiolaria). *Micropaleontology*, 23 (No. 3), 241-269.
- Petrushevskaya, M. G. 1967. Radiolarii otryadov Spumellaria i Nassellaria Antarkticheskoi oblasti (po materialam Sovetskogo, Antarkticheskogo ekspeditsii). *Issled. Fauny morei*, 3, 5-186.
- _____, 1971. On the natural system of Polycystine Radiolaria. *Proc. II Plankt. Conf. Rome 1970*, pp. 981-992.
- _____, 1975. Cenozoic radiolarians of the Antarctic, Leg 29. In Kennett, J. P., Houtz, R. E., et al., *Init. Repts. DSDP*, 29: Washington (U.S. Govt. Printing Office), 541-675.
- Petrushevskaya, M. G., and Kozlova, G., 1972. Radiolaria: Leg 14, Deep Sea Drilling Project. In Hayes, D. E., Pimm, A. C., et al., *Init. Repts. DSDP*, 14: Washington (U.S. Govt. Printing Office), 459-648.
- Reynolds, R. A., in prep. Neogene Radiolaria of the Northwest Pacific, eastern boundary current system [Ph.D. diss.]. Rice University, Houston.
- Riedel, W. R., 1957. Radiolaria: a preliminary stratigraphy. *Rept. Swed. Deep Sea Exped.*, 6 (No. 3), 61-96.
- _____, 1958. Radiolaria in Antarctic sediment. *B.A.N.Z. Antarctic Res. Exped. Rept.*, Series B, 6 (10), 217-255.
- _____, 1967. Subclass Radiolaria. In Harland, W. B., et al. (Eds.), *The Fossil Record*: London (Geological Society of London), pp. 291-298.
- _____, 1971. Systematic classification of Polycystine Radiolaria. In Funnell, B. M., and Riedel, W. R., (Eds.), *Micropalaeontology of the Oceans*: Cambridge (Cambridge University Press), pp. 649-661.
- Riedel, W. R., and Sanfilippo, A., Radiolaria, Leg 4, Deep Sea Drilling Project. In Bader, R., Gerard, R. D., et al., *Init. Repts. DSDP*, 4: Washington (U.S. Govt. Printing Office), 503-575.
- _____, 1971. Cenozoic Radiolaria from the western tropical Pacific, Leg 7. In Winterer, E. L., Riedel, W. R., et al., *Init. Repts. DSDP*, 7, Pt. 2: Washington (U.S. Govt. Printing Office), 1529-1672.
- _____, 1977. Cainozoic Radiolaria. In Ramsay, A. T. S. (Ed.), *Oceanic Micropalaeontology*: Cambridge (Cambridge University Press), pp. 847-912.
- _____, 1978. Stratigraphy and evolution of tropical Cenozoic radiolarians. *Micropaleontology*, 24 (No. 1), 61-96.
- Sanfilippo, A., and Riedel, W. R., 1970. Post-Eocene "closed" theoperid radiolarians. *Micropaleontology*, 16 (No. 4), 446-462.
- Westberg, M. J., and Riedel, W. R., 1978. Accuracy of radiolarian correlation in the Pacific Miocene. *Micropaleontology*, 24 (No. 1), 1-23.

PLATE 1

(For each figure the following information is given:
sample number, with intervals in cm; England Finder
coordinates; slide number; and maximum dimension in μm .)

- Figures 1-4 *Collosphaera* sp. A
 1. 438A-79-4, 102-104, Z 4/3, Sl. 1, 105.
 2. 438A-79-6, 102-104, Q 22/2, Sl. 2, 80 (end view).
 3. 440B-68-1, 21-22, S 37/2, Sl. 1, 95.
 4. 438A-8-1, 37-39, X 41/2, Sl. 1, 130.
- Figures 5-9 *Collosphaera pyloma* n. sp.
 5, 6. 438B-4-3, 37-40, U 1/3, Sl. 2, 105.
 7, 8. 438A-78-3, 90-92, H 14/3, Sl. 1, 125 (holotype).
 9. 438A-68, CC, 0 44/3, Sl. 2, 90 (end view).
- Figure 10 *Prunopyle titan* form A
 438A-73-2, 135-137, C 34/4, Sl. 1, 360.
- Figures 11-14 *Ommatartus* sp.
 11, 12. 438A-43-4, 49-51, M 16/3, Sl. 2, 200.
 13, 14. 438A-49-5, 44-46, Y 20/4, Sl. 1, 230.
- Figures 15, 16 *Clathrocyclas cabrilloensis*
 15. 438B-2, CC, J 7/1, Sl. 1, 170.
 16. 438A-45-1, 130-132, W 23/4, Sl. 2, 140.
- Figures 17, 18 *Lipmanella* sp. aff. *Theocorys redondoensis*
 17. 438A-30-4, 120-122, N 12/4, Sl. 2, 135.
 18. 438A-29-6, 140-142, D 83/1, Sl. 1, 200.
- Figure 19 *Lithocampe* sp.
 440B-56-3, 76-78, O 26/2, Sl. 1, 175.
- Figure 20 *Dictyophimus* sp. cf. *D. hirundo*
 438A-13-1, 27-29, G 37/4, Sl. 1, 90.
- Figures 21-22 *Lychnocanium* sp. cf. *L. grande*
 440-5-1, 123-125, V 20/2, Sl. 1, 165.
- Figures 23, 24 *Dictyophimus* sp.
 438A-51-4, 130-132, Q 23/3, Sl. 2, 145.

PLATE 1

